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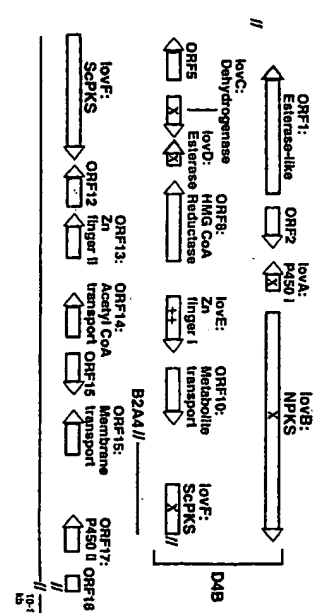
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(54) Title: METHOD OF PRODUCING ANTIHYPERCHOLESTEROLEMIC AGENTS

Lovastatin production genes



(57) Abstract:  
A method of increasing the production of lovastatin or monacolin J in a lovastatin-producing or non-lovastatin-producing organism is disclosed. In one embodiment, the method comprises the steps of transforming an organism with the A. terreus DAB segment, wherein the segment is translated and where an increase in lovastatin production occurs.

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## METHOD OF PRODUCING ANTIHYPERCHOLESTEROLEMIC AGENTS

## CROSS-REFERENCES TO RELATED APPLICATION

Not applicable.

STATEMENT REGARDING FEDERALLY SPONSORED  
RESEARCH AND DEVELOPMENT

5 This invention was made with United States government support awarded by the following agencies: NIH Grant No: AI43031. The United States has certain rights in this invention.

## BACKGROUND OF THE INVENTION

10 Cholesterol and other lipids are transported in body fluids by low-density lipoproteins (LDL) and high-density lipoproteins (HDL). Substances that effectuate mechanisms for lowering LDL-cholesterol may serve as effective antihypercholesterolemic agents because LDL levels are positively correlated with the risk of coronary artery disease.

15 MEVACOR (lovastatin; mevinolin) and ZOCOR (simvastatin) are members of a group of active antihypercholesterolemic agents that function by inhibiting the rate-limiting step in cellular cholesterol biosynthesis, namely the conversion of hydroxymethylglutarylcoenzyme A (HMG-CoA) into mevalonate by HMG-CoA reductase.

20 The general biosynthetic pathway of a naturally occurring HMG-CoA reductase inhibitor has been outlined by Moore, *et al.*, who showed that the biosynthesis of

mevinolin (lovastatin) by *Aspergillus terreus* ATCC 20542 begins with acetate and proceeds via a polyketide pathway (R.N. Moore, *et al.*, *J. Amer. Chem. Soc.* 107:3694-3701, 1985). Endo, *et al.* described similar biosynthetic

5 pathways in *Penicillium citrinum* NRRL 8082 and *Monascus ruber* M-4681 (A.Y. Endo, *et al.*, *J. Antibiot.* 38:444-448, 1985).

The recent commercial introduction of microbial HMG-CoA reductase inhibitors has fostered a need for high yielding production processes. Methods of improving process yield have included scaling up the process, improving the culture medium and simplifying the isolation.

15 Previous attempts to increase the biosynthesis of HMG-CoA reductase inhibitors at the level of gene expression have focused on increasing the concentration of triol polyketide synthase (TPKS), a multifunctional protein with at least six activities as evidenced by the product of the enzymatic activity (Moore, *supra*, 1985). TPKS is believed to be the rate-limiting enzymatic activity(ies) in the biosynthesis of the HMG-CoA reductase inhibitor compounds.

20 U.S. patent 5,744,350 identifies a DNA encoding triol polyketide synthase (TPKS) from *Aspergillus terreus*. "TPKS" is now preferred to TPKS as the acronym for "nonaketide polyketide synthase."

## SUMMARY OF THE INVENTION

In one embodiment, the present invention is a method of increasing the production of lovastatin in a lovastatin-producing organism. The method comprises the steps of transforming the organism with a nucleic acid sequence comprising the D4B segment, preferably comprising nucleotides 579 - 33,000 of SEQ ID NO:18 and 1 - 5,349 of SEQ ID NO:19. The nucleic acid sequence is transcribed and translated and an increase in lovastatin production occurs. Preferably, this increase is at least 2-fold.

In a preferred form of the present invention, the lovastatin-producing organism is selected from the group consisting of *A. terreus* ATCC 20542 and ATCC 20541.

In another embodiment, the method comprises the step of transforming the organism with the corresponding D4B segment isolated from a non-*A. terreus* lovastatin-producing organism.

In another embodiment, the present invention is a method of increasing the production of lovastatin in a lovastatin-producing organism, comprising the step of transforming the organism with the *lovE* gene, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.

In another embodiment of the present invention, one may increase the production of monacolin J in a non-lovastatin-producing organism comprising the steps of

transforming the organism with a nucleic acid sequence comprising the D4B segment. As a further step, one may additionally transform the organism with an entire *lovF* gene. If the entire *lovF* gene is added to the D4B segment, the organism will produce lovastatin.

In another embodiment, the present invention is the lovastatin production gene cluster, preferably SEQ ID NOs:18 and 19, and the individual genes comprising that cluster.

It is an object of the present invention to provide a method for increasing lovastatin and monacolin J production in both lovastatin-producing and non-lovastatin producing organisms.

Other objects, features and advantages of the present invention will become apparent after review of the specification, claims and drawings.

## DESCRIPTION OF DRAWINGS

Fig. 1 is a diagram of lovastatin production genes.

Fig. 2 is a schematic diagram of a hypothetical mevinolin/lovastatin biosynthesis pathway.

Fig. 3 is a comparative diagram of statins.

Fig. 4 is a schematic drawing of plasmid pMHM1264/CB24A.

Fig. 5 is a schematic drawing of plasmid pMHM1424.

Fig. 6 is a schematic drawing of plasmid CD4B/pMHM1263.

## DESCRIPTION OF THE INVENTION

In General

The Examples below disclose the cloning and sequencing of a cluster of 17 genes from *A. terreus* ATCC 20542, a strain that natively produces lovastatin (See Fig. 1). These genes flank the NPKS gene, which is known to be required for lovastatin production (see, for example, U.S. patent 5,744,350).

The DNA sequence of the cluster has been determined and is disclosed below at SEQ ID NOs:18 and 19.

Mutations in four of the genes (P450I/LovA, SEQ ID NO:22; dehydrogenase/LovC, SEQ ID NO:24; esterase/LovD, SEQ ID NO:25; and SCPKS/LovF, SEQ ID NO:29) have been isolated and demonstrate that each of these four individual genes is required for lovastatin production. These genes are indicated with an X symbol in Fig. 1 and referred to herein as the "*A. terreus* lovastatin gene cluster."

Another of the genes (Zn Finger I/LovE, SEQ ID NO:27) is thought to regulate the transcription of the other genes and causes a notable increase in lovastatin production when reintroduced into *A. terreus* ATCC 20542.

Applicants have used the following convention in naming the genes and proteins of the present invention. The genes and proteins are first named with either an "ORF" or "Lov" prefix and then named either numerically or alphabetically. "Lov" signifies genes shown to be essential for lovastatin production. Applicants have

also included a descriptor name that describes a probable function of the protein. For example, SEQ ID NO:1 is described as the "ORF1/esterase-like protein" because Applicants have compared the amino acid sequence to known esterases.

The portion of the gene cluster between ORF1/esterase-like protein and the mid-region of LovF/SCPKS is referred to as the "D4B segment". The *A. terreus* D4B segment is contained within a plasmid clone deposited as ATCC 98876. As described below, other lovastatin-producing organisms contain an analogous D4B segment comprising analogous genes. The present invention comprises a "D4B segment" isolated from other lovastatin-producing organisms. The arrangement of the genes within the D4B segment may be different in other organisms. We predict that the genes within these other segments will have at least 80% homology, at the nucleic acid level, with the genes disclosed herein. We envision that each of these lovastatin-producing organisms will comprise within their genomes a LovA, LovB, LovC, LovD, LovE and LovF gene.

We have determined that the D4B segment will confer production of monocolin J if the genes are all expressed, as we show below in an example using *A. nidulans*. We envision that adding the LovF gene to the D4B segment genes will result in the production of lovastatin in a non-lovastatin-producing organism.

Table 1, below, summarizes information regarding the

different protein and nucleic acid sequences of the present invention. SEQ ID NOS:1-17 are predicted translation products of various members of the gene cluster. SEQ ID NOS:18 and 19 are the entire DNA sequence of the gene cluster. SEQ ID NOS:21-36 are the genomic DNA sequences of the various members of the gene cluster and include the introns. These DNA sequences are reported in the Sequence Listing in the 5' - 3' orientation, although, as Fig. 1 indicates, some of these DNA sequences are in the inverted orientation in the actual cluster.

TABLE 1

SEQ ID NO.	DESCRIPTION	COMMENTS
SEQ ID NO: 1	Predicted amino acid sequence of ORF1/Esterase-like protein	Translation of 6 EXONS 6865-6568, 6462-5584, 5520-4822, 4774-3511, 3332-2372, 2301-1813 (reverse complement) FROM SEQ ID NO:18
SEQ ID NO: 2	Predicted amino acid sequence of ORF2	Translation of 1 EXON 7616-8602 FROM SEQ ID NO:18
SEQ ID NO: 3	Predicted amino acid sequence of LovA/P4501 protein	Translation of 1 EXON 10951-9980 (reverse complement) FROM SEQ ID NO:18
SEQ ID NO: 4	Predicted amino acid sequence of ORF3	Translation of 1 EXON 22760-21990 (reverse complement) FROM SEQ ID NO:18
SEQ ID NO: 5	Predicted amino acid sequence of LovC/Dehydrogenase	Translation of 3 EXONS 23158-23717, 23801-23912, 23991-24410 FROM SEQ ID NO:18
SEQ ID NO: 6	Predicted amino acid sequence of LovD/Esterase	Translation of 3 EXONS 26203-26080, 26005-25017, 24938-24810 (reverse complement) FROM SEQ ID NO:18

SEQ ID NO.	DESCRIPTION	COMMENTS
SEQ ID NO: 7	Predicted amino acid sequence of ORF8/HMG CoA Reductase	Translation of 5 EXONS 30062-29882, 29803-29745, 29664-27119, 27035-26779, 26722-26559 (reverse complement) FROM SEQ ID NO:18
SEQ ID NO: 8	Predicted amino acid sequence of LovE/Zn Finger 1	Translation of 1 EXON 31360-32871 FROM SEQ ID NO:18
SEQ ID NO: 9	Predicted amino acid sequence of ORF10/Metabolite transport	Translation of 8 EXONS 1400-1452, 1619-1695, 1770-1996, 2065-2088, 2154-2225, 2332-2865, 2939-3099, 3180-3560 FROM SEQ ID NO:19
SEQ ID NO: 10	Predicted amino acid sequence of LovF/ScPKS	Translation of 7 EXONS 4430-4627, 4709-4795, 4870-4927, 4985-5318, 5405-5912, 5986-6565, 6631-12464 FROM SEQ ID NO:19
SEQ ID NO: 11	Predicted amino acid sequence of ORF12	Translation of 3 EXONS 13596-13496, 13451-13063, 12968-12709 (reverse complement) FROM SEQ ID NO:19
SEQ ID NO: 12	Predicted amino acid sequence of ORF13/Zn Finger II	Translation of 5 EXONS 16608-16463, 16376-15572, 15519-15346, 15291-14825, 14767-14131 (reverse complement) FROM SEQ ID NO:19
SEQ ID NO: 13	Predicted amino acid sequence of ORF14/Aceyl CoA transport protein	Translation of 7 EXONS 19642-19571, 19502-19427, 19352-19227, 19158-19011, 18956-18663, 18587-18438, 18380-18341 (reverse complement) FROM SEQ ID NO:19
SEQ ID NO: 14	Predicted amino acid sequence of ORF15	Translation of 2 EXONS 20332-20574, 20631-21860 FROM SEQ ID NO:19
SEQ ID NO: 15	Predicted amino acid sequence of ORF16/Membrane transport protein	Translation of 5 EXONS 24521-24054, 23996-23936, 23876-23184, 23111-22977, 22924-22818 (reverse complement) FROM SEQ ID NO:19
SEQ ID NO: 16	Predicted amino acid sequence of ORF17/P45011 protein	Translation of 3 EXONS 28552-27673, 27606-27284, 27211-26837 (reverse complement) FROM SEQ ID NO:19

SEQ ID NO.	DESCRIPTION	COMMENTS
SEQ ID NO: 17	Predicted amino acid sequence of ORF18 (incomplete)	Translation of 2 EXONS 29826-30995, 31064-31328 (incomplete) FROM SEQ ID NO:19
SEQ ID NO: 18	DNA sequence of gene cluster- first 33,000 nucleotides	
SEQ ID NO: 19	DNA sequence of cluster- nucleotides 33,001-64,328 (renumbered 1-31,328)	
SEQ ID NO: 20	DNA sequence of ORF1/Esterase-like gene	Start = 6865 Stop = 1813 SEQ ID NO:18
SEQ ID NO: 21	DNA sequence of ORF2	Start = 7616 Stop = 8602 SEQ ID NO:18
SEQ ID NO: 22	DNA sequence of LovA/P450I gene	Start = 10951 Stop = 9980 SEQ ID NO:18
SEQ ID NO: 23	DNA sequence of ORF5	Start = 22760 Stop = 21990 SEQ ID NO:18
SEQ ID NO: 24	DNA sequence of LovC/Dhydrogenase	Start = 23158 Stop = 24410 SEQ ID NO:18
SEQ ID NO: 25	DNA sequence of LovD/Esterase	Start = 24810 Stop = 26203 SEQ ID NO:18
SEQ ID NO: 26	DNA sequence of ORF8/HMG CoA Reductase	Start = 30062 Stop = 26559 SEQ ID NO:18
SEQ ID NO: 27	DNA sequence of LovE/Zn Finger I	Start = 31360 Stop = 32871 SEQ ID NO:18
SEQ ID NO: 28	DNA sequence of ORF10/Metabolite transport	Start = 1400 Stop = 3560 SEQ ID NO:19
SEQ ID NO: 29	DNA sequence of LovF/ScPKS	Start = 4430 Stop = 12464 SEQ ID NO:19
SEQ ID NO: 30	DNA sequence of ORF12	Start = 13596 Stop = 12709 SEQ ID NO:19

SEQ ID NO.	DESCRIPTION	COMMENTS
SEQ ID NO: 31	DNA sequence of ORF13/Zn Finger II	Start = 16608 Stop = 14131 SEQ ID NO:19
SEQ ID NO: 32	DNA sequence of ORF14/Acetyl CoA transport gene	Start = 19642 Stop = 18341 SEQ ID NO:19
SEQ ID NO: 33	DNA sequence of ORF15	Start = 20332 Stop = 21860 SEQ ID NO:19
SEQ ID NO: 34	DNA sequence of ORF16/Membrane transport protein	Start = 24521 Stop = 22818 SEQ ID NO:19
SEQ ID NO: 35	DNA sequence of ORF17/P450II gene	Start = 28525 Stop = 26837 SEQ ID NO:19
SEQ ID NO: 36	DNA sequence of ORF18 (incomplete)	Start = 29826 to 31328 (incomplete) SEQ ID NO:19

Table 1 also notes the translation start and stop

points in the various gene sequences.

10 The sequence of the NPKS gene is not listed in SEQ ID NOS:21-36. This gene is characterized in U.S. patent 5,744,350. However, SEQ ID NOS:18 and 19 do contain the sequence of the NPKS gene within the context of the entire gene cluster.

15 To perform many embodiments of the present invention, one will need to recreate various genes or a portion of the gene cluster described herein. Applicants have provided sequence data in the Sequence Listing sufficient to allow one of skill in the art to construct numerous probes suitable to recreate the genes from an A. terreus genomic library. Applicants have also described below various methods for isolating A. terreus DNA.

Additionally, Applicants have deposited ATCC

Accession No. ATCC 98876, which contains clone pWHM1263 (CDB) and ATCC Accession No. ATCC 98877 which contains clone pWHM1265 (CB2A4). Both plasmids are described in more detail below. Fig. 4 describes clone

CB2A4/pWHM1265, and Fig. 6 describes clone CB4B/pWHM1263. Fig. 1 also indicates the boundaries of the D4B and B2A4 clones.

The clones and their inserts may be prepared from the ATCC deposits by methods known to those of skill in the art. The DNA from the clones may be isolated and any gene within the gene cluster may be isolated and utilized.

# 15 Increasing the Production of Lovastatin by Lovastatin-producing Fungi and Yeast

In one embodiment, the present invention is a method of increasing the production of lovastatin in a lovastatin-producing fungi and yeast, preferably *A. terreus* ATCC20542 and ATCC20541. Other examples of suitable lovastatin-producing fungi and yeast are listed in Table 2, below.

TABLE 2

Microorganisms other than <i>A. terreus</i> reported to produce lovastatin (mevastatin)	
Monascus (17 of 124 strains screened) species <sup>1</sup>	
5	<i>M. ruber</i>
	<i>M. purpureus</i>
	<i>M. pilosus</i>
	<i>M. vitreus</i>
	<i>M. pubigenus</i>
10	<i>Penicillium</i> sp. <sup>1</sup>
	<i>Hypomyces</i> sp.
	<i>Doratomyces</i> sp.
	<i>Phoma</i> sp.
15	<i>Eupentidium</i> sp.
	<i>Gymnosascus</i> sp.
	<i>Trichoderma</i> sp.
	<i>Pichia labensis</i> <sup>2</sup>
	<i>Candida carisioglycicola</i>
	<i>Aspergillus oryzae</i> <sup>3</sup>
20	<i>Doratomyces stemonitis</i>
	<i>Paeclomyces viroi</i>
	<i>Penicillium citrinum</i>
	<i>Penicillium chrysogenum</i>
	<i>Scopulariopsis brevicaulis</i>
25	<i>Trichoderma viride</i>
30	1. P. Jurek, I. Marinkova, V. Kren, Secondary Metabolites of the fungus <i>Monascus</i> : a review, <i>J. Ind. Microbiol.</i> 18:163-170 and references cited therein (1998).
	2. R. Gundlach, A. Penner, A. Charnan, A hydroxymethylglutaryl-CoA reductase inhibitor synthesized by yeast, <i>EBKS Microbiol. Lett.</i> 132:39-43 (1995).
	3. A.A. Shinde, Mevinolin production by some fungi, <i>Edin Microbiol.</i> 42:477-480 (1997).

By "increasing the production" we mean that the amount of lovastatin produced is increased by at least 2-fold, preferably by at least 5-fold. The examples below demonstrate two preferred methods for analyzing strains for lovastatin production. In method A, the spore suspension is inoculated into a flask of SEED medium and grown. The resulting seed culture is used to inoculate FM media and grown for six days. In fermentation method

B, one inoculates 50 ml of RPM media and grows this larger culture for 7 days.

Both cultures are extracted, pH adjusted, mixed with ethyl acetate and shaken for two hours. For analysis, 1 ml of the ethyl acetate layer is dried under a nitrogen stream and resuspended in methanol. For TLC analysis, a small amount of the extract is run on C18 reverse phase TLC plates in a solvent system of methanol; 0.1% phosphoric acid. The TLC plates are developed by spraying with phosphomolybdic acid in methanol and heating with a heat gun. The extracts are compared with authentic lovastatin, monacolin J, monacolin L and dihydromonoclon L.

If one wishes HPLC analysis, the examples below

describe the use of a Waters Nova-Pak C18 column used with a solvent system of acetonitrile and phosphoric acid. A Waters 996 Photodiode Array Detector will detect the metabolites. Lovastatin was detected at 238 nm.

In one embodiment, one would transform a lovastatin-producing fungi or yeast with the love/zinc finger I gene, preferably comprising the nucleotides of SEQ ID NO:27. The examples below predict that this will result in an increase of at least 5-7 fold. Preferably, the increase will be at least 2.0-fold.

One may also transform a lovastatin-producing fungi or yeast with the Love gene isolated from other lovastatin-producing fungi or yeast. One may obtain this

gene by use of a probe derived from SEQ ID NO:27 by methods known to those of skill in the art.

One may also transform lovastatin-producing fungi and yeast with the D4B segment of the lovastatin production gene cluster (see Fig. 1), preferably as found in ATCC accession number 98876. Alternatively, one may transform lovastatin-producing fungi or yeast with the entire gene cluster, as diagramed in Fig. 1.

We envision that to successfully increase lovastatin production, one may also wish to transform less than the entire gene cluster. Preferably, one may determine what the smallest possible segment is by deleting various portions of the gene cluster and determining whether lovastatin production is continually increased.

Similarly, if one begins with the D4B segment, one may delete various portions for the segment and determine whether lovastatin production is continually increased by at least 2-fold.

Modification of the LovB/NPKS gene would produce other HMG CoA inhibitors. For example, Fig. 3 diagrams the relationship between mevastatin, lovastatin, simvastatin and pravastatin. In one example, the methyl transferase domain of the NPKS gene may be replaced with an inactive form to make pravastatin. The HMG-CoA reductase inhibitors within this invention include, but are not limited to, compactin (ML-236B), lovastatin, simvastatin, pravastatin and mevastatin.



In another embodiment of the present invention, one may transform a lovastatin-producing organism with the genes described above and obtain the production of an HMG CoA reductase inhibitor with a structure different from monacolin J, monacolin L or lovastatin. Alterations in the side chain attached to C8 are the most likely possibility but other alterations may occur. These alterations would happen through the native biochemistry of the organism.

10 If one wishes to express the *A. terreus* genes in yeast, one may wish to consult examples in which others have engineered fungal secondary metabolism genes for expression in yeast. (See for example, J. T. Kealey, et al., *Proc. Natl. Acad. Sci. USA* 95:505-509 (1998)). The exact approach could be used with the NPKS (LovB) and ScPKS (LovF) genes, and a somewhat simpler approach with the other lovastatin genes in their cDNA forms.

Production of HMG-CoA Reductase Inhibitors by Fungi and Yeast That Do Not Natively Produce Inhibitors.

20 In another embodiment, the present invention is the production of HMG-CoA reductase inhibitors, such as lovastatin, by fungi and yeast that do not natively produce lovastatin. An example of a suitable fungi or yeast is *A. nidulans* and *S. cerevisiae*, respectively.

25 For this embodiment one preferably transforms the genes within the D4B segment into the non-inhibitor-producing strain. By this method, one would produce

monacolin J (See Fig. 2) which could be chemically converted to lovastatin by one of skill in the art.

Monacolin J, in its lactone form obtained by treatment with anhydrous acid under dehydrative conditions, is preferably treated with a derivative of (2S)-2-methylbutyric acid, in which the carboxyl group has been suitable activated for undergoing esterification, and the resulting lovastatin is isolated by conventional methods. For example, see WO 33538, U.S. patent 4,444,784 and *J. Med. Chem.* 29:849 (1986). These are citations for synthesis of simvastatin from monacolin J. One would use the same method, but use the (2S)-2-methylbutyrate derivative to make lovastatin.

15 In another embodiment of the present invention, one would transform the genes within the D4B segment, including an entire LovF/ScPKS gene, into the non-inhibitor-producing organism. By this method, one would produce lovastatin in a non-lovastatin-producing organism.

20 In another embodiment of the present invention, one may transform a non-lovastatin-producing organism with the genes described above and obtain the production of an HMG CoA reductase inhibitor with a structure different from monacolin J, monacolin L or lovastatin, as described above.

25 Modification of the LovB/NPKS gene would produce other inhibitors. For example, Fig. 3 diagrams the relationship between mevastatin, lovastatin, simvastatin

and pravastatin. In one example, the methyl transferase domain of the NPKS gene may be replaced with an inactive form to make pravastatin. The HMG-CoA reductase

inhibitors within this invention include, but are not

5 limited to, compactin (ML-236B), lovastatin, simvastatin, pravastatin and mevastatin.

#### Production of Intermediate Materials

In another embodiment, the present invention is a method of isolating intermediate materials in the production of lovastatin and analogs such as mevastatin and simvastatin. For example, the Examples below demonstrate the disruption of the lovastatin projection gene cluster with mutagenized LovC, LovD, LovF, LovA or LovB genes. Disruption of many of these genetic elements of the lovastatin production gene cluster will result in accumulation of intermediate materials. Therefore, to practice this embodiment of the present invention, one would transform a suitable lovastatin-producing host with a mutagenized gene within the D4B segment, as described below.

Many other mutations would be suitable to destroy the function of LovC, LovD, LovF, LovA or LovB. All that is necessary is these genes be disrupted to the extent that they are non-functional.

#### Production of Lovastatin Analogs

In another embodiment, the present invention provides a method for engineering the production of

lovastatin analogs in such organisms as fungi or yeast, using monacolin J as the starting point.

#### Isolated DNA Segments

In another embodiment, the present invention is a DNA segment capable of conferring lovastatin or monacolin J production or increase in lovastatin or monacolin J production in yeast or fungi. In a preferred example, this segment is the "D4B segment" that is deposited at ATCC 98876. The nucleotide sequence of this segment is found in residues 579 - 33,000 of SEQ ID NO:18 and residues 1 - 5,349 of SEQ ID NO:19.

In another embodiment, the present invention is the entire *A. terreus* lovastatin gene cluster, as exemplified by SEQ ID NOs:18 and 19 and ATCC deposits 98876 and 98877.

The present invention is also the individual genes that make up the *A. terreus* lovastatin gene cluster.

Therefore, the present invention is a nucleic acid segment selected from the group of consisting of SEQ ID NOs:20 - 36. Preferably, the present invention is the coding region found within SEQ ID NOs:20 - 36 and described in Table 1. The present invention is also a mutagenized version of SEQ ID NOs:22, 24, 25 and 29, wherein the gene is mutagenized to be non-functional in terms of lovastatin or monacolin J production.

### Organisms with Increased Lovastatin or Monocolin J Production

In another embodiment, the present invention are the organisms described above. These organisms include lovastatin-producing organisms, preferably yeast and fungi, that have been engineered to display at least a 2-fold increase in lovastatin or monocolin J production. The organisms also include non-lovastatin-producing organisms, preferably yeast or fungi, that have been engineered to produce monocolin J or lovastatin.

### Antifungal Compounds

Applicants note that lovastatin, monocolin J, monocolin L and dihydromonocolin L all have varying degrees of antifungal activity. Applicants envision that the present invention is also useful for providing antifungal compounds and organisms engineered to express antifungal compounds. Preferably, one would measure the antifungal properties of a compound in the manner of N. Lomovskaya, et al., *Microbiology* 143:875-883, 1997.

Measurement of inhibition of yeast growth can be found in R. Ikeura, et al., *J. Antibiotics* 41:1148, 1988. The same general methods could be used for all fungi. Both of these references are hereby incorporated by reference.

### 1. General Methods and Procedures

#### Construction of an *A. terreus* ATCC20542 genomic library.

*A. terreus* ATCC20542 genomic DNA was partially digested with *Sau3AI* so as to produce an average fragment size of 40 - 50 kb. The partially digested genomic DNA was then separated on a sucrose gradient and the 40 - 50 kb fraction was collected. Cosmid AN26 (Taylor and Borgmann, *Emgal Genet. Newsletter* 43, 1996) was prepared by digestion with *ClaI*, dephosphorylated with CIP, then digested with *BamHI* to create the two cosmid arms. Ligation reactions with genomic DNA fragments and cosmid arms were optimized and packaged using Gigapack III XL packaging extract (Stratagene). The packaged cosmid library was infected into *E. coli* JM109 and plated out onto LB agar (Sambrook, et al., Molecular Cloning—A Laboratory Manual, 2nd ed. Cold Spring Harbour Laboratory Press, 1989; other standard methods used can be found here also) with ampicillin (50 µg/ml) plates. After checking for the presence of insert DNA in a selection of clones, 5000 colonies were picked into LB plus 50 µg/ml ampicillin filled microtitre plates and grown overnight at 37°C. The colonies were replica plated onto nylon membranes (Amersham Hybond-N). Glycerol was added at a final concentration of 15% (v/v) to the microtitre plates and these were stored at -70°C.

PCT/US99/29583  
Isolation of genomic clones containing the lovastatin biosynthesis cluster.

A 2.8 kb *EcoRI* fragment from pTPKS100 containing part of the NPKS gene (Vinci, et al., U.S. Patent No.

5 5,744,350) was gel-isolated and labelled with digoxigenin using the Genius Kit II (Boehringer Mannheim). This labelled fragment was hybridized (65°C, 5x SSC) with the nylon membranes containing the *A. terreus* genomic

library, then washed (65°C, 0.1x SSC). Two positive clones were identified, pWHM1263 (cD4B) and pWHM1264

(cJ3A). Two of these clones, pWHM1263 (cD4B) and

pWHM1265 (cB2A4), have been deposited in the ATCC

(American Type Culture Collection, 10801 University Boulevard, Menassas, VA 20110) at accession number ATCC

15 98876 and 98877, respectively, under the terms and

conditions of the Budapest Treaty. The presence of the NPKS gene was confirmed initially by restriction digestion and later by DNA sequencing.

Overlapping clones were found by repeating the

20 hybridization process using labelled fragments from both ends of the insert in pWHM1263. This resulted in the

isolation of pWHM1265-1270 (cB2A4, cL3E2, cJ3B5, cO2B5,

cR3B2, cW3B1) from downstream of the NPKS gene and

pWHM1271 (cQ1F1) from upstream of NPKS. All these clones

25 were transformed into *E. coli* strain STBL2 (Stratagene) to help prevent rearrangements.

Fig. 4 is a diagram of the cB2A4/pWHM1265 clone.

This clone contains an insert of approximately 43 kb in

AN26 and includes the nucleotide sequence from at least nucleotides 4988 of SEQ ID NO:19 to nucleotide 31,328 of SEQ ID NO:19 and 10 - 14 kb of uncharacterized DNA. Fig. 6 is a schematic diagram of cD4B/pWHM1263. This clone

5 contains a 37,770 bp insert in AN26 and contains

nucleotides 579 - 33,000 of SEQ ID NO:18 and nucleotides

1 - 5,349 of SEQ ID NO:19.

#### Sequencing strategy and analysis.

A series of overlapping subclones (pWHM1272-

10 pWHM1415) were constructed in pSPORT1 (Gibco-BRL) and

pGEM3 (Promega). Plasmid DNAs for sequencing were

prepared using the QiaPrep spin miniprep kit (Qiagen).

Cycle sequencing was carried out using the AmpliTaq FS or

BigDye reagents (ABI) and were analyzed using a ABI model

15 373 or 377 DNA Sequencer. Primer walking was performed

by synthesis of 18-22 bp oligonucleotide primers based on the sequenced DNA strand, with the help of the Oligo 4.05

program (National Biosciences, Inc.). Every region of

DNA was sequenced at least once on both strands. Direct

20 sequencing of cosmids and PCR products was used to

confirm adjoining regions where no overlapping clones

existed. DNA sequence analysis and manipulations were

performed using SeqMan (DNASTAR) and SeqEd (ABI)

software. Assignments of putative ORFs, including

25 putative introns, were performed with the aid of BLAST

2.0 searches (Atschul, et al., *Nucl. Acids Res.* 25:3389-

3402, 1997), and the Genetics Computer Group (GCG)

programs (Program Manual for the Wisconsin Package,

Version 8, September 1994, Genetics Computer Group,

Madison, WI), version 8.1.

Isolation and characterization of *lovF* (ScPKS, ORF11),  
*lovD* (EST1, ORF7), *lovC* (DH, ORF6), and *lovA* (P4501,  
ORF3) mutants.

#### *lovF*

To disrupt the polyketide synthase gene, *lovF*, a 1.7  
kb *EcoRI* fragment internal to the *lovF* gene was subcloned  
from pWHM1265 into pSPORT1 to give pWHM1291. The ScPKS  
fragment was then subcloned from this vector, as an  
Acc65I - *HindIII* fragment, into pPLOA (Vinci, et al.,  
U.S. Patent No. 5,744,350) to give pWHM1416. This vector  
contains the phleomycin (Zeocin, obtained from  
Invitrogen) resistance gene for selection in *A. terreus*.

*A. terreus* ATCC20542 was then transformed to Zeocin  
resistance with this plasmid as described below.

Transformants were screened for lovastatin production as  
described below (Method A). In one of the transformants,  
WMH1731, lovastatin production was abolished and a new

compound accumulated. This new compound comigrated with  
monacolin J on TLC and HPLC according to the methods  
described below. Semi-preparative HPLC was used to  
partially purify the major product which was then  
analyzed by HPLC - MS. The same mass and fragmentation  
pattern as authentic monacolin J was observed. To  
confirm the disruption of the *lovF* gene, total genomic  
DNA was prepared from wild-type *A. terreus* ATCC20542 and  
the WMH1731 mutant strain. The genomic DNA was digested

with *BamHI* and *HindIII*, electrophoresed on an agarose gel  
and capillary blotted onto a nylon membrane. The  
membrane was hybridized with the 1.7 kb *EcoRI* fragment  
from pWHM1416 labelled using the Genius II kit

(Boehringer Mannheim) using the conditions described  
previously. The wild-type strain had hybridizing bands  
at 4.2 kb for *BamHI* and 11.5 kb for *HindIII*. As  
predicted, the WMH1731 mutant strain had hybridizing  
bands at 6.5 kb and 2.2 kb for *BamHI* and 11 kb and 7.8 kb  
for *HindIII* confirming the homologous integration of a  
single copy of pWHM1416 at the *lovF* locus.

#### *lovD*

To disrupt the putative esterase/carboxypeptidase-  
like gene, *lovD*, a 4.8 kb *NciI* - *EcoRI* fragment from  
pWHM1263 was subcloned into pSPORT1 to give pWHM1274.

This plasmid was digested with *HindIII* and *BsiWI* and a  
1.8 kb fragment was isolated. The plasmid was also  
digested with *HindIII* and *BamHI* and the 6.6 kb fragment  
was isolated. pPLOA was digested with *BamHI* and Acc65I  
and the 2.1 kb fragment containing the phleomycin  
resistance marker was purified. These three fragments  
were ligated together and used to transform competent *E.*  
*coli* cells. The expected plasmid, pWHM1417, containing  
the phleomycin resistance gene flanked by the beginning  
and the end of the *lovD* gene was isolated. This plasmid  
was linearized by digestion with *XbaI* or *RsaI* before

being used to transform *A. terreus* ATCC20542 to Zeocin resistance. Transformants were screened for lovastatin production as described below (Method A). In one of the transformants, WMH1732, lovastatin production was abolished and a new compound accumulated. This new compound comigrated with monacolin J on TLC and HPLC according to the methods described below. Semi-preparative HPLC was used to partially purify the major product which was then analyzed by HPLC - MS. The same mass and fragmentation pattern as authentic monacolin J was observed. To confirm the disruption of the *lovD* gene, total genomic DNA was prepared from wild type *A. terreus* ATCC20542 and the WMH1732 mutant strain. The genomic DNA was digested with *ApaI*, run out on an agarose gel and capillary blotted onto a nylon membrane. The membrane was hybridized with the 4.8 kb *NotI* - *EcoRI* fragment from pWHM1274 labelled using the Genius II kit using the conditions described previously. The wild-type strain had hybridizing bands at 9 kb, 8.4 kb and 1.5 kb. As predicted the mutant strain had hybridizing bands at 9 kb, 8 kb, 3 kb and 1.5 kb confirming the homologous integration of a single copy of pWHM1417 at the *lovD* locus.

# lovA

To disrupt the cytochrome P450 I gene, *lovA*, an 11 kb *Acc65I* - *EcoRI* fragment from pWHM1263 was subcloned into pGEM3 to give pWHM1272. From this plasmid a 2.1 kb

*ApaI* - *SnaBI* fragment was purified and ligated to *ApaI* - *EcoRV* digested pPLOA to give p450Phleo (pWHM1418). From this plasmid a 4.2 kb *ApaI* - *NotI* fragment was purified and ligated with a 1.8 kb *EagI* - *KpnI* fragment from pWHM1272 and *ApaI* - *KpnI* digested pGEM7 to give p450Dphleo (pWHM1419) which contains the *lovA* gene disrupted by the phleomycin resistance gene. This plasmid was then digested with *KpnI* and *ApaI* and the resulting fragment was used to transform *A. terreus* ATCC20542 to Zeocin resistance. Transformants were screened for lovastatin production as described below (Method A). In one of the transformants, WMH1733, lovastatin production was abolished and two new compounds accumulated. Genomic DNA was prepared from this strain and from *A. terreus* ATCC20542, digested with *EagI*, run out on an agarose gel, and capillary blotted onto a nylon membrane. The membrane was hybridized with the 6 kb *ApaI* - *KpnI* fragment from pWHM1419 labelled using the Genius II kit using the conditions described previously. The wild-type strain had hybridizing bands at 2.0 kb, 1.9 kb and 1.1 kb. Mutant strain WMH1733 had hybridizing bands at 2.5 kb, 2.0 kb, 1.1 kb and 0.7 kb confirming the homologous integration of a single copy of the fragment from pWHM1419 at the *lovA* locus.

**lovC**

To disrupt the dehydrogenase-like gene, *lovC*, a 2 kb *EcoRI* - *BglII* fragment from pTPKS100 was ligated with a 1.7 kb *EcoRI* - *SacI* fragment from pWHM1274 and *BglII* - *SacI* digested litmus 28 (New England Biolabs) to produce pDH1 (pWHM1420). Another plasmid pDH2 (pWHM1421) was constructed from a 2.2 kb *Acc65I* - *SacI* fragment from pWHM1274, a 2.1 kb *HindIII* - *SacI* fragment from pPLOA containing the phleomycin resistance gene and *HindIII* - *Acc65I* digested litmus 28. The disruption vector pDH-dis (pWHM1422) was constructed by ligating together a 2.5 kb *BglII* - *HpaI* fragment from pWHM1420, a 4.3 kb *EcoRV* - *KpnI* fragment from pWHM1421 and *BglII* - *KpnI* digested litmus 28. This plasmid was digested with *BglII* and *KpnI* and the resulting 6.8 kb fragment was used to transform *A. terreus* ATCC20542 to Zeocin resistance. Transformants were screened for lovastatin production as described below (Method A). In two of the transformants, WMH1734 and WMH1735, lovastatin production was abolished.

Genomic DNA was prepared from these strains and from *A. terreus* ATCC20542, digested with *EagI*, run out on an agarose gel, and capillary blotted onto a nylon membrane. The membrane was hybridized with the 6.8 kb *BglII* - *KpnI* fragment from pWHM1422 labelled using the Genius II kit using the conditions described previously. The wild type strain had hybridizing bands at 5 kb, 1.5 kb and 1.3 kb.

Mutant strain WMH1734 had hybridizing bands at 4.9 kb, 1.3 kb, 1.0 kb and 0.7 kb confirming the homologous integration of a single copy of the fragment from pWHM1422 at the *lovC* locus. The other mutant strain, WMH1735, had a similar banding pattern but with additional hybridizing bands indicating that multiple integration events had occurred, one of which was at the *lovC* locus.

Construction and characterization of the *A. terreus* strain with extra copies of *lovC*.

A 10.4 kb *NotI* - *EcoRI* fragment containing the putative regulatory gene, *lovE* was subcloned from pWHM1263 to pSPORT1 to give pWHM1276. From this plasmid a 3.9 kb *HindIII* - *BamHI* fragment was subcloned into pGEM7 to give pWHM1423. The regulatory gene was subcloned from this vector into pPLOA as an *SacI* - *XbaRI* fragment to give pWHM1424 (Fig. 5). pWHM1424 contains nucleotides 30,055 - 33,000 from SEQ ID NO:18 and nucleotides 1 - 1,026 from SEQ ID NO:19.

Extra copies of the regulatory gene were introduced into *A. terreus* ATCC20542 by transformation to Zeocin resistance with pWHM1424. Transformants were fermented (method A) and screened for lovastatin production initially by TLC analysis. Most of the transformants appeared to be producing significantly more lovastatin than the wild-type strain. The yields of lovastatin from the two transformant strains, WMH1736 and WMH1737, which had the most elevated levels compared to the wild-type

was quantified by HPLC as described below. These were found to produce 7-fold and 5-fold more lovastatin than the *A. terreus* ATCC20542 strain.

Because of the way that the DNA integrates

5 (ectopically), each transformant is or can be unique, genotypically and phenotypically. However, some will be overproducers; others may exhibit no difference, for unknown reasons.

#### 10 Heterologous expression of the lovastatin biosynthesis genes.

To place the NPKS gene (*lovB*) under the control of the inducible *alcA* promoter, the 11.5 kb *KpnI* - *AvrII* fragment from pTPKS100 containing the NPKS open reading frame was ligated into pAL3 (Waring, et al., Gene 79:119,

15 1989) previously digested with *KpnI* and *XbaI*. The resulting plasmid was designated pAL3TPKS (WHM1425). The polymerase chain reaction was used to amplify the NPKS gene sequence between the NPKS promoter region just upstream of the translational start codon and a *AgeI* site

20 internal to NPKS. The design of the forward primer introduced a *KpnI* site 31 bases from the translational start codon allowing the NPKS to be placed against the *alcA* promoter but also incorporating upstream elements

from the *A. terreus* system. Amplification was performed 25 using Vent DNA polymerase with pTPKS100 as template and 1  $\mu$ mol of each primer in a final volume of 100  $\mu$ l using the manufacturer's buffer recommendations. After an initial

denaturation cycle of 10 minutes at 95°C amplification was achieved with 30 cycles of 95°C for 1 minute; 55°C for 1 minute and 72°C for 1.5 minutes. The final cycle was followed by 10 minutes at 72°C to ensure complete

5 polymerization. The amplified product (1.7 kb) was digested with *KpnI* and *AgeI* and ligated into pWHM1425 that had been digested with the same enzymes and gel isolated. The resulting plasmid was designated

10 pAL3TPKSNT (pWHM1426). The region introduced by PCR was sequenced on a ABI automated DNA sequencer to ensure sequence fidelity. This plasmid was then used to transform *A. nidulans* strain A722 (Fungal Genetics Stock Centre) to uridine prototrophy.

Transformants were grown by inoculating 0.5 ml of 15 spore suspension ( $10^8$  c.f.u./ml) in 50 ml YEPD in a 250 ml un baffled flask. This was then grown for 20 hours at 250 rpm and 37°C (New Brunswick Scientific Series 25

20 Incubator Shaker). The mycelia were then harvested by filtration through Miracloth (Calbiochem), rinsed with sterile, distilled water, and inoculated into fresh 250 ml un baffled flasks containing 50 ml AMM + lactose + 10 mM cyclopentanone and grown for a further 20 hours under the same conditions. The mycelia were harvested by

25 filtration using Miracloth (Calbiochem), squeezed as dry as possible and frozen in liquid nitrogen. Protein extracts for SDS-PAGE and western analysis were prepared as described in Kennedy and Turner, *Molec. Gen. Genet.* (1996), 253:189-197, 1996.



One transformant, WMH1738, was shown to have a large protein (>200 kDa) visible on a SDS-PAGE gel that cross reacted with the affinity purified NPKS antibodies (Parilabs). This strain WMH1738 was transformed to hygromycin B resistance with pMHM1263. Transformant colonies were screened for lovastatin resistance and for the production of new metabolites as described below and two strains WMH1739 and WMH1740 were chosen for further analysis. Both of these strains were found to be significantly resistant (up to 100 µg/ml on solid media) to lovastatin compared with the host strain. This was analyzed by streaking 10 µl of a spore suspension on solid AMM plates containing lovastatin at 0, 0.1, 0.5, 1, 5, 10, 50 and 100 µg/ml and incubating at 37°C. Strains WMH1739 and WMH1740 were compared to strains WMH1741 and WMH1742 which were derivatives of WMH1738 transformed to hygromycin resistance with AM26. Strains WMH1739 and -1740 exhibited no inhibition of growth at any of these lovastatin concentrations whereas strains WMH1741 and -1742 showed slight inhibition of growth at 5 µg/ml and almost complete growth inhibition at 50 µg/ml. The two lovastatin resistant strains were fermented in lovastatin-producing conditions using fermentation method B and extracts were analyzed for lovastatin related metabolites as described below. Both strains were found to produce new metabolites. One compound that was common to both comigrated with monacolin J on TLC and HPLC analysis by the methods described below. Semi-

preparative HPLC was used to partially purify some of this compound, which was then analyzed by HPLC - MS. It had the same mass and fragmentation pattern as authentic monacolin J. The other compound, found in only one of the strains, comigrated with monacolin L on TLC and HPLC.

#### METHODS

##### Solid medium for growth of *A. terreus*

For the generation of spore suspensions *A. terreus* strains were grown on CM agar at 30°C for 4 to 5 days.

CM Agar (for CM liquid medium the agar was omitted):  
50 ml Clutterbuck's salts (Vinci, et al., U.S.

Patent No. 5,744,350)

2 ml Vogel's trace elements (Vinci, et al., U.S. Patent No. 5,744,350)

0.5% Difco Bacto tryptone  
0.5% Difco Bacto yeast extract

1% glucose  
2% Difco Bacto agar

in 1 liter of distilled water

##### Clutterbuck's salts:

12% NaNO<sub>3</sub>

1.02% KCl

1.04% MgSO<sub>4</sub>·7H<sub>2</sub>O

25

3.04% KH<sub>2</sub>PO<sub>4</sub>

##### Vogel's trace elements:

0.004% ZnCl<sub>2</sub>

0.02% FeCl<sub>3</sub>

30

0.001% CuCl<sub>2</sub>

0.001% MnCl<sub>2</sub>·4H<sub>2</sub>O

0.001% Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub>·10H<sub>2</sub>O

0.001% (NH<sub>4</sub>)<sub>2</sub>MoO<sub>7</sub>·7H<sub>2</sub>O

For long term storage *A. terreus* spores were suspended in SSS (10% glycerol, 5% lactose) and stored at -70°C.

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For the generation of spore stocks *A. nidulans* was grown on the following solid growth medium (ACM) for 3 to 4 days at 37°C.

ACM:

2% Difco Bacto malt extract  
0.1% Difco Bacto peptone  
2% glucose  
2% agar (Difco, Detroit, MI)

For strains which required para-aminobenzoic acid

10 (PABA) for growth, PABA was added to a final

concentration of 1 µg/ml. For strains which required

uracil and uridine these were added at 20 mM and 10 mM,

respectively. Spores were suspended in Tween 80 - saline solution (0.025% Tween 80, 0.8% NaCl) and stored at 4°C.

15 AMM:

0.6% (w/v) NaNO<sub>3</sub>  
0.052% (w/v) KCl  
0.152% (w/v) KH<sub>2</sub>PO<sub>4</sub>  
0.052% (w/v) MgSO<sub>4</sub>·7H<sub>2</sub>O  
1% (w/v) glucose  
0.1% (v/v) AMM trace elements solution  
pH to 6.5 and make up to 1 liter with distilled water.

20

For preparation of plates 2% (w/v) Difco Bacto agar

25 was added. If required the glucose can be omitted and an alternative carbon source (e.g., lactose added at the same concentration). For the preparation of transformation plates KCl was added at 4.47% (w/v) (0.6 M).

30 AMM trace elements solution:

0.1% (w/v) FeSO<sub>4</sub>·7H<sub>2</sub>O  
0.88% (w/v) ZnSO<sub>4</sub>·7H<sub>2</sub>O  
0.04% (w/v) CuSO<sub>4</sub>·5H<sub>2</sub>O  
0.015% (w/v) MnSO<sub>4</sub>·4H<sub>2</sub>O  
0.01% (w/v) Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub>·10H<sub>2</sub>O

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0.005% (NH<sub>4</sub>)<sub>2</sub>MoO<sub>4</sub>·7H<sub>2</sub>O  
distilled water to 1 liter

Large scale genomic DNA preparation from *A. terreus* for genomic library construction.

5 A 2.5 ml aliquot of spore suspension (10<sup>8</sup> c.f.u./ml)

was used to inoculate 500 ml of liquid CM medium and

grown for 20 hours at 30°C and 200 rpm. The mycelium was

harvested by filtration through Miracloth (Calbiochem)

and rinsed extensively with water then TSE [150 mM NaCl,

10 100 mM Na<sub>2</sub>EDTA, 50 mM Tris-HCl pH 8.0]. The mycelium was

squeezed dry, broken into small pellets and frozen in

liquid nitrogen then ground to a fine powder in a pre-

chilled pestle and mortar followed by transfer to a

500 ml flask. Fifty ml of extraction buffer [150 mM

15 NaCl, 100 mM Na<sub>2</sub>EDTA, 50 mM Tris-HCl pH 8.0, 2% (w/v) SDS]

and 10 ml of toluene was added to the flask which was

shaken at 60 rpm for 72 hours. This mixture was

centrifuged at 1000 x g for 15 minutes and the

supernatant was removed and extracted with an equal

20 volume of chloroform:isoamyl alcohol (24:1 vol/vol).

This mixture was centrifuged at 10,000 x g for 30 minutes

at 15°C. The aqueous layer was carefully removed and 1.1

volumes of ethanol was layered on top. The DNA was

spooled out from the resulting suspension and resuspended

25 in 5 ml TE [10 mM Tris-HCl pH 8.0, 1 mM EDTA] + 50 µg/ml

RNase and 100 µg/ml proteinase K then incubated at 37°C

for 2 hours. The mixture was extracted again with

chloroform:isoamyl alcohol (24:1) and the DNA was spooled

out as before. Following resuspension in 1 ml of TE the

DNA was extracted once with phenol:chloroform:isoamyl alcohol (25:24:1, vol/vol), once with chloroform:isoamyl alcohol (24:1) and precipitated with 0.6 volumes isopropanol. The DNA clot was removed, dried briefly and resuspended in 0.5 ml TE.

**Small scale genomic DNA preparation from *A. terreus* for Southern blot.**

A 0.5 ml aliquot of spore suspension ( $10^8$  c.f.u./ml) was used to inoculate 100 ml of liquid CM and grown for 20 hours at 30°C and 200 rpm. The mycelium was harvested by filtration through Miracloth (Calbiochem) and rinsed extensively with water then TSE (150 mM NaCl, 100 mM Na<sub>2</sub>EDTA, 50 mM Tris-HCl pH 8.0). The mycelium was squeezed dry, broken into small pellets and frozen in liquid nitrogen. The mycelium was ground to a fine powder in a pre-chilled pestle and mortar and transferred to a mortar pre-heated to 65°C. Three ml of lysis buffer (0.5 M NaCl, 10 mM Tris-HCl pH 7.5, 10 mM EDTA, 1% (w/v) SDS) at 65°C was added and 0.3 ml of 10% (w/v) cetyltrimethylammonium bromide in 0.7 M NaCl. After thorough mixing to form a slurry, 3 ml of phenol:chloroform:isoamyl alcohol (25:24:1) was added. This mixture was transferred to a Corex tube and incubated at 65°C for 15 minutes. Following centrifugation at 12,000 x g for 15 minutes at 4°C the aqueous phase was carefully removed and re-extracted once with phenol, once with phenol:chloroform:isoamyl alcohol (25:24:1) and once with chloroform:isoamyl alcohol (24:1). The DNA was precipitated from the extract by

addition of 0.1 volume of 3 M sodium acetate pH 5 and 0.6 volumes isopropanol then collected by centrifugation (10,000 x g, 10 minutes, 4°C). After washing with 70% ethanol the pellet was briefly dried and resuspended in TE + RNase (50 µg/ml).

**Transformation of *A. terreus*.**

A 0.5 ml aliquot of spore suspension ( $10^8$  c.f.u./ml) was used to inoculate 100 ml of liquid CM and grown for 20 hours at 30°C and 200 rpm. The mycelium was harvested by centrifugation at 2000 x g for 15 minutes at 4°C and washed twice with an aqueous solution containing 0.27 M CaCl<sub>2</sub> and 0.6 M NaCl. To produce protoplasts the washed mycelia was resuspended in 20 ml of the same solution containing 5 mg/ml Novozym 234 (NovoNordisk) and incubated at 30°C for 1 - 3 hours with gentle agitation. Protoplasts were separated from undigested mycelia by filtration through Miracloth (Calbiochem). The protoplast suspension was diluted with an equal volume of STC1700 (1.2 M sorbitol, 10 mM Tris-HCl pH 7.5, 35 mM NaCl) and incubated on ice for 10 minutes. The protoplasts were collected by centrifugation (2000 x g, 10 minutes, 4°C), washed with STC1700 and resuspended in 1 ml STC1700. Plasmid DNA, purified using Qiagen columns, (2 - 5 µg in 10 µl) was added to 150 µl of protoplast suspension and incubated at room temperature for 25 minutes. PEG solution (60% (w/v) polyethylene glycol 4000, 50 mM CaCl<sub>2</sub>, 10 mM Tris-HCl pH 7.5) was added to the DNA/protoplasts mixture in three steps: 250 µl,

250  $\mu$ l, and 850  $\mu$ l with mixing after each addition. The suspension was incubated at room temperature for 25 minutes then diluted to 10 ml with STC1700. Protoplasts were collected by centrifugation as above and diluted with 500  $\mu$ l STC1700. 100  $\mu$ l aliquots of this mixture were plated onto osmotically stabilized plates [CM medium containing 3% (w/v) Difco Bacto agar and 23.4% (w/v) mannitol, 15 ml of agar per plate]. After 4 hours growth at 30°C, 25 ml of OL agar [1% (w/v) Difco Bacto peptone, 1% (w/v) Difco Bacto agar, 200  $\mu$ g/ml Zeocin] was overlaid onto each dish. The plates were incubated for 3 - 4 days at 30°C before transformant colonies were picked. These were streaked to single colonies twice on selective media (CM + 100  $\mu$ g/ml Zeocin) before spore suspensions were prepared.

#### Transformation of *A. nidulans*.

A 0.5 ml aliquot of spore suspension ( $10^8$  c.f.u./ml) was used to inoculate 100 ml of YEPD [2% (w/v) Difco Bacto yeast extract, 2% (w/v) glucose, 0.1% Difco Bacto peptone] liquid medium including necessary supplements and grown for 20 hours at 37°C and 200 rpm. The mycelia was harvested by centrifugation (2000 x g, 10 minutes, 4°C) and washed twice with 0.6 M KCl. To generate protoplasts the mycelia was resuspended in 20 ml of 0.6 M KCl containing 5 mg/ml Novozym 234 and incubated at 30°C for 1 - 2 hours with gentle shaking. Protoplasts were separated from undigested mycelia by filtration through Miracloth (Calbiochem). The protoplasts were harvested

by centrifugation as described above and washed twice with 0.6 M KCl, then resuspended in 10 ml 0.6 M KCl + 50 mM CaCl<sub>2</sub>. After counting in a haemocytometer the protoplasts were harvested by centrifugation as before and resuspended to a final concentration of  $5 \times 10^6$  protoplasts/ml. To 50  $\mu$ l of protoplast suspension, 5  $\mu$ l of DNA (2 - 5  $\mu$ g, purified using Qiagen columns) was added, then 12.5  $\mu$ l of PEG solution [25% (w/v) PEG 6000, 50 mM CaCl<sub>2</sub>, 10 mM Tris - HCl pH 7.5] and the mixture was incubated on ice for 20 minutes. A further 0.5 ml of PEG solution was added and the mixture was incubated on ice for a further 5 minutes. A 1 ml aliquot of 0.6 M KCl + 50 mM CaCl<sub>2</sub> was added and the protoplasts were plated out in 50  $\mu$ l, 200  $\mu$ l, and 400  $\mu$ l aliquots. For transformation to uridine prototrophy, protoplasts were plated out onto AMM + 0.6 M KCl plates without adding uridine or uracil supplements. Plates were incubated at 37°C for 3 - 4 days when transformants were picked. For transformation to hygromycin B resistance protoplasts were plated out onto AMM + 0.6 M KCl plates (15 ml) and incubated for 4 hours at 30°C. 30 ml of 1% peptone, 1% agar, 1 mg/ml hygromycin B was then used to overlay the plates, which were incubated for 3 - 4 days when transformants were picked. Transformants from both methods were streaked out to single colonies on selective media (i.e., lacking uridine/uracil supplements or containing 1  $\mu$ g/ml hygromycin B) twice before spore suspensions were made.

## Analysis of strains for lovastatin production.

Two fermentation methods were used for the analysis of lovastatin production. In Method A, 0.5 ml of spore suspension (10<sup>8</sup> c.f.u./ml) was inoculated into 25 ml of SEED medium in 250 ml unbaffled flasks and grown for 18 hours at 250 rpm and 30°C (New Brunswick Scientific Model 25 incubator/shaker). A 1 ml portion of the resulting seed culture was used to inoculate 25 ml of FM in a 250 ml unbaffled flask and grown for 6 days in the conditions described above. Fermentation Method B involved inoculating 50 ml of RPM in a 250 ml unbaffled flask with 0.5 ml of spore suspension (10<sup>8</sup> c.f.u./ml) and growing at 30°C and 250 rpm for 7 days in a New Brunswick Scientific Series 25 Incubator Shaker.

## 15 SEED medium:

- 0.5% (w/v) Sigma corn steep liquor
- 4% (w/v) tomato paste
- 1% (w/v) oat flour
- 1% (w/v) glucose
- 1% (v/v) Vogel's trace elements
- distilled water to 1 l

## FM:

- 4.5% (w/v) glucose
- 2.4% (w/v) Sigma peptonized milk
- 0.25% (w/v) Difco Bacto yeast extract
- 0.25% (w/v) polyethylene glycol 2000
- distilled water up to 1 l

## RPM:

- 4% (w/v) lactose
- 0.3% (w/v) rapeseed meal
- 0.2% (w/v) KNO<sub>3</sub>
- 0.3% (w/v) KH<sub>2</sub>PO<sub>4</sub>
- 0.05% (w/v) MgSO<sub>4</sub> · 7H<sub>2</sub>O
- 0.05% (w/v) NaCl
- 0.05% (v/v) Sigma anti-foam B
- 0.05% (v/v) trace elements solution
- pH to 6.5 and made up to 1 l with distilled water.

## Trace elements solution is:

- 0.16% (w/v) MnSO<sub>4</sub>
- 0.34% (w/v) ZnSO<sub>4</sub> · 7H<sub>2</sub>O
- 0.2% (w/v) CoCl<sub>2</sub> · 6H<sub>2</sub>O
- 0.5% (w/v) FeSO<sub>4</sub> · 7H<sub>2</sub>O

made up to 1 liter with distilled water.

The cultures were extracted by adjusting the pH of the media to 3 with HCl, adding an equal volume of ethyl acetate, and shaking the mixture on a New Brunswick Scientific Series 25 incubator/shaker at 250 rpm for 2 hours. For analysis, 1 ml of the ethyl acetate layer was dried under a nitrogen stream and resuspended in 0.1 ml of methanol. For TLC analysis 10 µl of this extract was run on C-18 reverse phase TLC plates (RP-18 F<sub>100</sub> - Merck) in a solvent system of methanol:0.1% phosphoric acid (9:1). TLC plates were developed by spraying with 10% phosphomolybdic acid in methanol and heating with a heat gun. Extracts were compared with authentic lovastatin, monacolin J, monacolin L, and dityromonacolin L (acid and lactone forms). For HPLC analysis a Waters Nova-Pak C<sub>18</sub> (3.9 x 150 mm) column was used with a solvent system of acetonitrile (B) and 0.1% phosphoric acid (A). The column was eluted with a preprogrammed gradient of 0 to 100% B into A over 25 minutes using gradient 7 (Waters Millennium Software) with a flow rate of 1.5 ml/min and metabolites were detected with a Waters 996 Photodiode Array Detector; lovastatin was detected at 238 nm. For purification of metabolites a Waters Prep Nova-Pak HR C<sub>18</sub> (7.8 x 300 mm) column was used. The same solvent system as above was used with gradient of 0 to 100% B in A over

75 minutes at a flow rate of 4.5 ml/min. Fractions were collected manually, back extracted with ethyl acetate and dried. For HPLC-MS an Aquapore OD-300 7 micron (1.0 x 100 mm) column was used with a gradient of 0 to 100% acetonitrile into A (0.05% TFA) over 30 minutes at a flow rate of 0.02 ml/min.

5

## CLAIMS

We claim:

1. A method of increasing the production of lovastatin in a lovastatin-producing organism, comprising the steps of transforming the organism with the D4B segment, wherein the segment is transcribed and translated, and wherein an increase in lovastatin production occurs.
2. The method of claim 1 wherein the D4B segment is the *A. terreus* D4B segment.
3. The method of claim 1, wherein the D4B segment is identical to nucleotides 579 - 33,000 of SEQ ID NO:18 and 1 - 5,349 of SEQ ID NO:19.
4. The method of claim 1, wherein the lovastatin-producing organism is selected from the group consisting of *A. terreus* ATCC 20542 and ATCC 20541.
5. The method of claim 1, wherein the organism is selected from the group consisting of fungi and yeast.
6. The method of claim 1 wherein the increase is at least 2-fold.

7. The method of claim 1 wherein the nucleic acid sequence is identical to a sequence isolated from ATCC 98876.

8. The method of claim 1 additionally comprising transforming the organism with the entire *A. terreus* lovastatin gene cluster.

9. The method of claim 8 wherein the gene cluster comprises SEQ ID Nos:18 and 19.

10. The method of claim 8 wherein the nucleic acid sequence of the gene cluster is identical to sequences isolated from ATCC 98876 and 98877.

11. A method of increasing the production of monacolin J in a lovastatin-producing organism, comprising the steps of transforming the organism with the DAB segment, wherein the segment is translated, and wherein an increase monacolin J production occurs.

12. A method of increasing the production of lovastatin in a lovastatin-producing organism, comprising the step of transforming the organism with the LOVE gene, wherein the nucleic acid sequence is translated, and wherein an increase in lovastatin production occurs.

13. The method of claim 12 wherein the increase is at least 2.0-fold.

14. The method of claim 13 wherein the increase is at least 5-fold.

15. The method of claim 12 wherein the nucleotide sequence of the LOVE gene comprises SEQ ID NO:27.

16. A method of increasing the production of lovastatin in a lovastatin-producing organism comprising the steps of transforming the organism with a nucleic acid sequence comprising a truncated version of the *A. terreus* DAB segment, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.

17. A method of increasing the production of lovastatin in a lovastatin-producing organism comprising the steps of transforming the organism with a nucleic acid sequence comprising a truncated version of the *A. terreus* lovastatin-producing gene cluster, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.

18. A method of increasing or conferring the production of monacolin J in a non-lovastatin-producing organism comprising the steps of transforming the organism with a nucleic acid sequence comprising the D4B segment, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in monacolin J production occurs.

5

19. The method of claim 18 wherein the D4B segment is the *A. terreus* D4B segment.

20. The method of claim 18 wherein the D4B segment comprises nucleotides 579 - 33,000 of SEQ ID NO:18 and 1-5,349 of SEQ ID NO:19.

21. The method of claim 18 additionally comprising the step of converting the monacolin J into lovastatin.

22. The method of claim 18 additionally comprising the step of transforming the organism with a nucleic acid sequence comprising the *lovF* gene, wherein the nucleic acid sequence is transcribed and translated and wherein an increase in lovastatin production occurs.

5

23. An isolated nucleic acid sequence selected from the group consisting of SEQ ID NOS:20 - 36.

24. A lovastatin-producing organism, wherein the organism has been genetically modified to have increased lovastatin production, wherein the increase is at least 2-fold.

25. The organism of claim 24, wherein the organism is a yeast or a fungi.

26. A non-lovastatin producing organism, wherein the organism has been genetically modified to produce monacolin J.

27. The organism of claim 26, wherein the organism is a yeast or a fungi.

28. A non-lovastatin producing organism, wherein the organism has been genetically modified to produce lovastatin.

29. The organism of claim 28 wherein the organism is a yeast or a fungi.



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FIG. 2

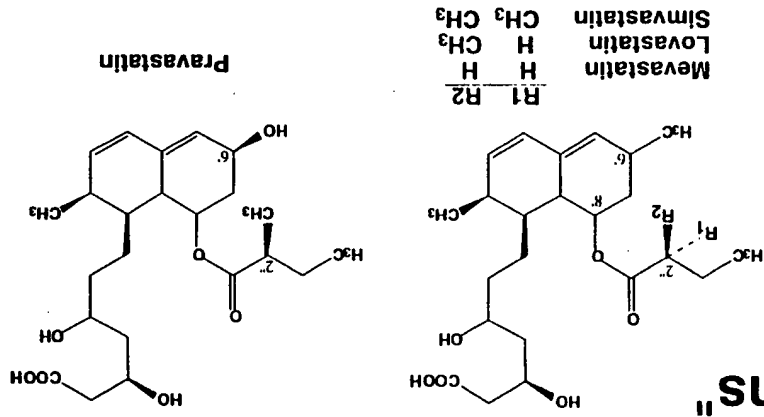


FIG. 3

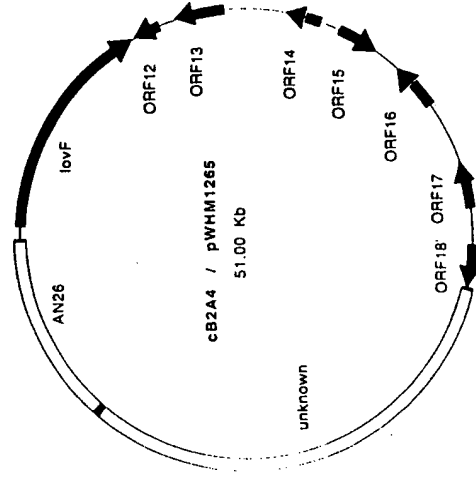
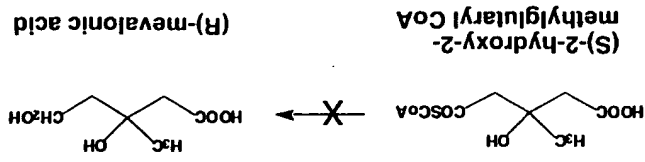


FIG. 4

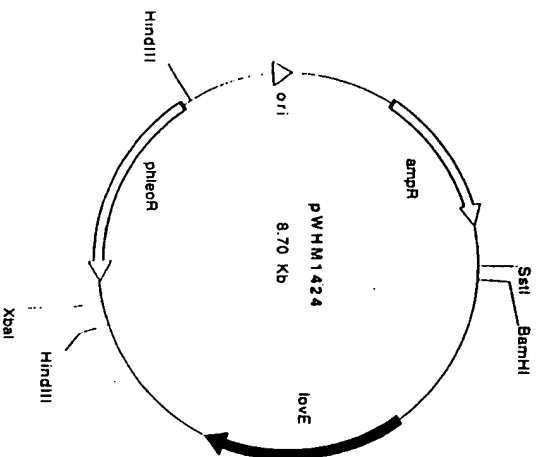


FIG. 5

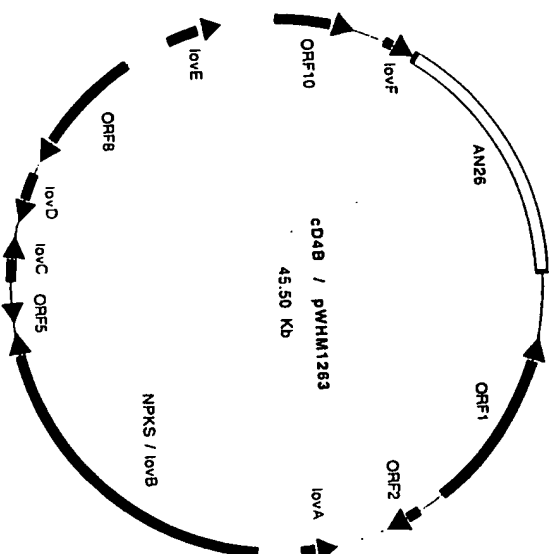


FIG. 6

## SEQUENCE LISTING

<110> Wisconsin Alumni Research Foundation  
Hutchinson, Charles R.  
Kennedy, Jonathan R.M.I.  
Park, Cheonseck R.M.I.

<120> METHOD OF PRODUCING ANTIHYPERCHOLESTEROLEMIC AGENTS

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<170> PatentIn Ver. 2.0

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35 40 45

Ala Lys Arg Asp Ala Leu Lys Val Pro Leu Arg Ile Leu Pro Leu  
50 55 60

Gly Ala Ser Ile Thr Trp Gly Tyr Leu Ser Ser Thr Gly Asn Gly Tyr  
65 70 75 80

Arg Lys Pro Leu Arg Asp Lys Leu Arg Phe Glu Gly Trp Glu Val Asp  
85 90 95

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100 105 110

Ala Ala Ala Asn Ser Leu Ala Tyr Lys Pro Asn Val Val Leu Ile Asn  
115 120 125

Ala Gly Thr Asn Asp Cys Asp Tyr Asn Val Asp Pro Ala Asn Ala Gly  
130 135 140

Glu Arg Met Arg Ser Leu Ile Glu Thr Leu Ile Gly Ala Pro Asp Met  
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Ala Asn Thr Leu Ile Val Leu Ser Thr Leu Ile Pro Ser Gly Ser Thr  
165 170 175

Thr Leu Glu Ala Asn Arg Pro Ser Val Asn Ala Gln Phe Arg Glu Leu  
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Val Leu Asp Met Arg Glu Ala Gln Asn Val Ser Ile Val Leu Ala Asp  
195 200 205

Met Asp Pro Pro Ala Pro Ser Pro Gly Asn Asn Trp Ile Thr Tyr Pro  
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Asp Asn Phe Ala Asp Asn Lys His Pro Asn Asp Tyr Gly Tyr Ser Gln  
225 230 235 240

Met Ala Asp Ile Trp Tyr Asn Ala Ile Tyr Asn Ala Ala Val Ala Glu  
245 250 255

Leu Ile Val Lys Pro Ala Asp Leu Asp Ile Ser Ser Thr Gly Thr Cys  
260 265 270

Asp Lys Glu Tyr Gly Ser Gly Val Tyr Ala Gly Phe Thr Gln Gln  
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Gly Ser Gly Glu Asp Asp Gly Ile Tyr Arg His Asp Ser Glu Tyr Ser  
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Gly Ala Leu Phe Thr Val Arg Ala Gly Lys Gly Ala Ala Asp Pro Tyr  
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Lys Asp Asp Asp Glu Leu His Phe Phe Gly Arg Leu Tyr Thr Arg  
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260 265 270  
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Gln Thr His Arg Asn Ser Ala Asp Gly Arg Leu Arg Tyr Asp Asp Ser  
245 250 255  
Val Tyr Phe Arg Ala Asp Gly Glu Cys Phe Gly Gly Gln Gly Val  
260 265 270  
Phe Ser Gly Pro Gly Ser Tyr Met Lys Val Leu His Ser Leu Leu Lys  
275 280 285  
Arg Asp Gly Leu Leu Leu Gln Pro Gln Thr Val Asp Leu Met Phe Gln  
290 295 300  
Pro Ala Leu Glu Pro Arg Leu Glu Glu Gln Met Asn Gln His Met Asp  
305 310 315 320  
Ala Ser Pro His Ile Asn Tyr Gly Gly Pro Met Pro Met Val Leu Arg  
325 330 335  
Arg Ser Phe Gly Leu Gly Gly Ile Ile Ala Leu Glu Asp Leu Asp Gly  
340 345 350  
Glu Asn Trp Arg Arg Lys Gly Ser Leu Thr Phe Gly Gly Gly Pro Asn  
355 360 365  
Ile Val Trp Gln Ile Asp Pro Lys Ala Gly Leu Cys Thr Leu Ala Phe  
370 375 380  
Phe Gln Leu Glu Pro Trp Asn Asp Pro Val Cys Arg Asp Leu Thr Arg  
385 390 395 400



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Thr Phe Glu His Ala Ile Tyr Ala Gln Tyr Gln Gln Gly  
405 410

<210> 7  
<211> 1068  
<212> PRT  
<213> Aspergillus terreus

<400> 7  
Met Asp Pro Val Val Arg Lys Pro Asp Pro Gly Gly Val Gln His Arg  
1 5 10 15  
Val Thr Lys Ala Leu Arg Ala Ile Val Gly His Ala Cys Arg His Pro  
20 25 30  
Ile His Thr Leu Leu Val Thr Ala Leu Thr Ala Ala Thr Thr His Leu  
35 40 45  
His Val Leu Glu Gly Thr Tyr Gln Ala Thr His Arg Glu Ala Ser Ala  
50 55 60  
Tyr Lys Tyr Gln Ile Asp Asp Arg Pro Lys Val Pro Glu Asp Gly Gln  
65 70 75  
Ser Asp Phe His Tyr Ala Leu Val Thr Leu Asp Leu Pro Gly Ala Ser  
85 90 95  
Val Asp Ala Ser Ile Pro Phe Leu Ser Asn Thr Leu Ser Gly Phe Leu  
100 105 110  
Gly Ala Glu Gln Thr Thr Pro Thr Pro Asp Ser Ser Pro Ser Pro Asp  
115 120 125  
His Ser Ala Leu Thr Phe Arg Val Pro Tyr Ser Ser Gln Leu Asp Gly Phe  
130 135 140  
Leu Gln Ala Val Glu Ile Ile Pro Ser Glu Lys Glu Asp Asp Ser Tyr  
145 150 155  
Arg Leu Arg Ser Pro Arg Glu Glu Gly Ser Pro Arg Ser Leu Gly His  
160 165 170  
Tyr Leu Gly Ser Ser Tyr Leu Ser Phe Leu His Arg Val His His Ala  
180 185 190  
Glu Thr Val Asp Leu Val Ile Ile Gly Leu Ser Tyr Leu Ala Met Asn  
195 200 205  
Met Thr Val Val Ser Leu Phe Arg Val Met Arg His Leu Gly Ser Arg  
210 215 220  
Phe Tyr Leu Ala Ala Ser Val Leu Leu Ser Gly Ala Phe Ala Phe Val  
225 230 235  
Leu Gly Leu Gly Ile Thr Thr Thr Cys Asp Val Pro Val Asp Met Leu  
240 245 250  
Leu Leu Phe Glu Gly Ile Pro Tyr Leu Val Leu Thr Val Gly Phe Glu  
255 260 265  
Lys Pro Ile Gln Leu Thr Arg Ala Val Leu Cys Val Ser Glu Glu Leu  
270 275 280  
Tyr Gly Gly Gly Gln Arg Gln Val Pro Asn Gly Ala Ser Ser Asp Asp  
285 290 295 300

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Ser Arg Gln Asn Gln Leu Ile Pro Asn Ile Ile Gln Leu Ala Val Asp  
305 310 315 320  
Arg Glu Gly Tyr Ile Val Arg Ser Tyr Leu Leu Glu Ile Gly Ala  
325 330 335  
Leu Ala Leu Gly Ala Val Leu Arg Pro Lys Asp Ser Leu Gly His Phe  
340 345 350  
Cys Phe Leu Ala Ala Tyr Thr Leu Leu Ile Asp Ala Val Leu Leu Phe  
355 360 365  
Thr Phe Tyr Ala Thr Ile Leu Cys Val Lys Leu Glu Ile Thr Arg Ile  
370 375 380  
Arg Ser Pro Gly Gly Leu Gly Gln Val Asn Ala Lys His Pro Ser Gly  
385 390 395  
Ile Phe Gly His Lys Val Lys Ser Thr Asn Ile Thr Tyr Tyr Lys Leu  
400 405 410  
Leu Thr Val Gly Gly Phe Val Leu Cys His Phe Leu Gln Leu Ser Pro  
415 420 425  
Phe Tyr Arg Val Met Gly Glu Tyr Met Ala Asn Gly Thr Leu Pro  
430 435 440  
Pro Thr Ala Val Ser Pro Phe Lys Glu Ala Ala Asn Gly Leu Asn Glu  
445 450 455  
Ile Tyr Leu Thr Ala Arg Val Glu Gly Phe Glu Thr Arg Val Thr Val  
460 465 470  
Leu Pro Pro Leu Gln Tyr Val Leu Glu Ser Ala Gly Phe Asn Ile Ser  
475 480 485  
Ala Thr Lys Arg Ser Thr Phe Asp Gly Val Leu Asp Gly Leu Glu Ser  
490 495 500  
Pro Leu Gly Arg Leu Cys Leu Met Gly Ala Leu Val Val Ser Leu Val  
505 510 515  
Leu Asn Asn His Leu Ile His Ala Ala Arg Tyr His Ala Tyr Pro Gln  
520 525 530  
Ala Arg Glu Ser Ala Val Pro Asp Gly Ser Tyr Leu Ser Val Pro Cys  
535 540 545  
Ser Ala Thr Ala Pro Glu Val Cys Thr Arg Pro Pro Glu Glu Thr Glu  
550 555 560  
Ala Leu Leu Lys Ser Asn Gln Ala Ala Ser Leu Thr Asp Asp Glu Leu  
565 570 575  
Val Glu Leu Cys Leu Arg Gly Lys Ile Ala Gly Tyr Ser Leu Glu Lys  
580 585 590  
Thr Leu Glu Arg Ile Ala Ala Gly Ser Ser Arg Ser Val Thr Arg Leu  
595 600 605  
Glu Ala Phe Thr Arg Ala Val Arg Ile Arg Arg Ala Ala Val Ser Lys  
610 615 620  
625 630 635 640  
Thr Pro Ser Thr Gln Asn Leu Cys Ser Gly Leu Ala Glu Ser Leu Leu  
645 650 655

12

Pro Tyr Arg Asp Tyr Asn Tyr Glu Leu Val His Gly Ala Cys Cys Glu  
 660 665  
 Asn Val Val Gly Tyr Leu Pro Leu Pro Leu Gly Val Ala Gly Pro Met  
 675 680 685  
 Val Ile Asp Gly Gln Ala Leu Phe Ile Pro Met Ala Thr Thr Glu Gly  
 690 695 700  
 Val Leu Val Ala Ser Ala Ser Arg Gly Cys Lys Ala Ile Asn Ala Gly  
 705 710 715 720 725  
 Gly Gly Ala Thr Thr Met Leu Lys Gly Asp Gly Met Thr Arg Gly Pro  
 725 730 735  
 Cys Leu Arg Phe Pro Ser Ala Gln Arg Ala Ala Glu Ala Gln Arg Tyr  
 740 745 750  
 Val Glu Ser Pro Leu Gly His Glu Val Leu Ala Ala Phe Asn Ala  
 755 760 765  
 Thr Ser Arg Phe Ala Arg Leu Gln Thr Leu Thr Val Ala Gln Ala Gly  
 770 775 780  
 Ile Tyr Leu Tyr Ile Arg Phe Arg Thr Thr Thr Gly Asp Ala Met Gly  
 785 790 795 800  
 Met Asn Met Ile Ser Lys Gly Val Glu Lys Ala Leu Glu Ala Met Ala  
 805 810 815  
 Ala Glu Gly Gly Phe Pro Asp Met His Thr Val Thr Leu Ser Gly Asn  
 820 825 830  
 Phe Cys Ser Asp Lys Lys Ser Ala Ala Ile Asn Trp Ile Gly Gly Arg  
 835 840 845  
 Gly Lys Ser Val Ile Ala Glu Ala Thr Ile Pro Ala Glu Thr Val Arg  
 850 855 860  
 Gln Val Leu Lys Thr Asp Val Asp Ala Leu Val Glu Leu Asn Thr Ala  
 865 870 875  
 Lys Asn Leu Val Gly Ser Ala Met Ala Gly Ser Leu Gly Gly Phe Asn  
 885 890 895  
 Ala His Ala Ser Asn Leu Val Gln Ala Val Phe Leu Ala Thr Gly Gln  
 900 905 910  
 Asp Pro Ala Gln Asn Val Glu Ser Ser Ser Cys Ile Thr Thr Met Lys  
 915 920 925  
 Asn Ile Asp Gly Asn Leu His Ile Ala Val Ser Met Pro Ser Met Glu  
 930 935 940  
 Val Gly Thr Ile Gly Gly Gly Thr Ile Leu Glu Ala Gln Gly Ala Met  
 945 950 955 960  
 Leu Asp Leu Leu Gly Val Arg Gly Ala His Ser Thr Glu Pro Gly Ala  
 965 970 975  
 Asn Ala Arg Arg Leu Ala Arg Ile Val Ala Ala Val Leu Ala Gly  
 980 985 990  
 Glu Leu Ser Thr Cys Ala Ala Leu Ala Ala Gly His Leu Val Asn Ala  
 995 1000

His Met Gln His Asn Arg Thr Ser Lys Asp Ala Ile Ser Gly Thr Glu  
 1010 1015 1020  
 Tyr Gly Ala Ile Arg Thr Pro Val Tyr Val Val Ile Leu Gln His Ala  
 1025 1030 1035 1040  
 Gly Asp Ile His Phe Val Gln Ile Glu Tyr Lys Asn Thr Tyr Leu Arg  
 1045 1050 1055  
 Arg Lys Val Pro Thr Leu Ser Cys Asn Leu Gly Arg  
 1060  
 <210> 8  
 <211> 503  
 <212> PRT  
 <213> Aspergillus terreus  
 <400> 8  
 1 Met Ala Ala Asp Gln Gly Ile Phe Thr Asn Ser Val Thr Leu Ser Pro  
 5 10 15  
 Val Glu Gly Ser Arg Thr Gly Gly Thr Leu Pro Arg Arg Ala Phe Arg  
 20 25 30  
 Arg Ser Cys Asp Arg Cys His Ala Gln Lys Ile Lys Cys Thr Gly Asn  
 35 40 45  
 Lys Glu Val Thr Gly Arg Ala Pro Cys Gln Arg Cys Gln Gln Ala Gly  
 50 55 60  
 Leu Arg Cys Val Tyr Ser Glu Arg Cys Pro Lys Arg Lys Leu Arg Gln  
 65 70 75 80  
 Ser Arg Ala Ala Asp Leu Val Ser Ala Asp Pro Asp Pro Cys Leu His  
 85 90 95  
 Met Ser Ser Pro Pro Val Pro Ser Gln Ser Leu Pro Leu Asp Val Ser  
 100 105 110  
 Glu Ser His Ser Ser Asn Thr Ser Arg Gln Phe Leu Asp Pro Pro Asp  
 115 120 125  
 Ser Tyr Asp Trp Ser Trp Thr Ser Ile Gly Thr Asp Glu Ala Ile Asp  
 130 135 140  
 Thr Asp Cys Trp Gly Leu Ser Gln Cys Asp Gly Gly Phe Ser Cys Gln  
 145 150 155 160  
 Leu Glu Pro Thr Leu Pro Asp Leu Ser Pro Ser Pro Phe Glu Ser Thr Val  
 165 170 175  
 Glu Lys Ala Pro Leu Pro Pro Val Ser Ser Asp Ile Ala Arg Ala Ala  
 180 185 190  
 Ser Ala Gln Arg Glu Leu Phe Asp Asp Leu Ser Ala Val Ser Gln Glu  
 195 200 205  
 Leu Glu Glu Ile Leu Leu Ala Val Thr Val Glu Trp Pro Lys Gln Glu  
 210 215 220  
 Ile Trp Thr Arg Ala Ser Pro His Ser Pro Thr Ala Ser Arg Glu Arg  
 225 230 235 240  
 Ile Ala Gln Arg Arg Gln Asn Val Trp Ala Asn Trp Leu Thr Asp Leu  
 245 250 255

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His Met Ser Leu Asp Pro Ile Gly Met Phe Asn Ala Ser Arg  
260 265 270  
Arg Leu Leu Thr Val Leu Arg Gln Gln Ala Asp Cys His Gln  
275 280 285  
Gly Thr Leu Asp Gln Cys Leu Arg Thr Lys Asn Leu Phe Thr Ala Val  
290 295 300  
His Cys Tyr Ile Leu Asn Val Arg Ile Leu Thr Ala Ile Ser Gln Leu  
305 310 315 320  
Leu Leu Ser Gln Ile Arg Arg Thr Gln Asn Ser His Met Ser Pro Leu  
325 330 335  
Glu Gly Ser Arg Ser Gln Ser Pro Ser Arg Asp Thr Ser Ser Ser  
340 345 350  
Ser Gly His Ser Ser Val Asp Thr Ile Pro Phe Phe Ser Gln Asn Leu  
355 360 365  
Pro Ile Gly Gln Leu Phe Ser Tyr Val Asp Pro Leu Thr His Ala Leu  
370 375 380  
Phe Ser Ala Cys Thr Thr Leu His Val Gly Val Gln Leu Arg Gln  
385 390 395 400  
Asn Gln Ile Thr Leu Gly Val His Ser Ala Gln Gly Ile Ala Ala Ser  
405 410 415  
Ile Ser Met Ser Gly Gln Pro Gly Gln Asp Ile Ala Arg Thr Gly Ala  
420 425 430  
Thr Asn Ser Ala Arg Cys Gln Gln Gln Pro Thr Thr Pro Ala Ala Arg  
435 440 445  
Val Leu Phe Met Phe Leu Ser Asp Gln Gly Ala Phe Gln Gln Ala Lys  
450 455 460  
Ser Ala Gly Ser Arg Gly Arg Thr Ile Ala Ala Leu Arg Arg Cys Tyr  
465 470 475 480  
Glu Asp Ile Phe Ser Leu Ala Arg Lys His Lys His Gly Met Leu Arg  
485 490 495  
Asp Leu Asn Asn Ile Pro Pro  
500

<210> 9  
<211> 542  
<212> PRT  
<213> Aspergillus terreus

<400> 9  
Met Thr Ser His His Gly Gln Thr Gln Lys Pro Gln Ser Asn Thr Ala  
1 5 10 15  
Gln Met Gln Ile Asn His Val Thr Gly Leu Arg Leu Gly Leu Val Val  
20 25 30  
Val Ser Val Thr Leu Val Ala Phe Leu Met Leu Asp Met Ser Ile  
35 40 45  
Ile Val Thr Ala Ile Pro His Ile Thr Ala Gln Phe His Ser Leu Gly  
50 55 60

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Asp Val Gly Trp Tyr Gly Ser Ala Tyr Leu Leu Ser Ser Cys Ala Leu  
65 70 75 80  
Gln Pro Leu Ala Gly Lys Leu Tyr Thr Leu Leu Thr Leu Lys Tyr Thr  
85 90 95  
Phe Leu Ala Phe Leu Gly Leu Phe Gln Ile Gly Ser Val Leu Cys Gly  
100 105 110  
Thr Ala Arg Ser Ser Thr Met Leu Ile Val Gly Arg Ala Val Ala Gly  
115 120 125  
Met Gly Gly Ser Gly Leu Thr Asn Gly Ala Ile Thr Ile Leu Ser Ala  
130 135 140  
Ala Ala Pro Lys Gln Gln Gln Pro Leu Leu Ile Gly Ile Met Met Gly  
145 150 155 160  
Leu Ser Gln Ile Ala Ile Val Cys Gly Pro Leu Leu Gly Gly Ala Phe  
165 170 175  
Thr Gln His Ala Ser Trp Arg Trp Cys Phe Tyr Ile Asn Leu Pro Ile  
180 185 190  
Gly Ala Phe Ala Thr Phe Leu Leu Leu Val Ile Gln Ile Pro Asn Arg  
195 200 205  
Leu Pro Ser Thr Ser Asp Ser Thr Thr Asp Gly Thr Asn Pro Lys Arg  
210 215 220  
Arg Gly Ala Arg Asp Val Leu Thr Gln Leu Asp Phe Leu Gly Phe Val  
225 230 235 240  
Leu Phe Ala Gly Phe Ala Ile Met Ile Ser Leu Ala Leu Gln Trp Gly  
245 250 255  
Gly Ser Asp Tyr Ala Trp Asn Ser Ser Val Ile Ile Gly Leu Phe Cys  
260 265 270  
Ala Ala Gly Val Ser Leu Val Leu Phe Gly Cys Trp Gln Arg Val  
275 280 285  
Gly Gly Ala Val Ala Met Ile Pro Ile Ser Val Ala Ser Arg Arg Gln  
290 295 300  
Val Trp Cys Ser Cys Phe Phe Leu Gly Phe Phe Ser Gly Ala Leu Leu  
305 310 315 320  
Ile Phe Ser Tyr Tyr Leu Pro Ile Tyr Phe Gln Ala Val Lys Asn Val  
325 330 335  
Ser Pro Thr Met Ser Gly Val Tyr Met Leu Pro Gly Ile Gly Gln  
340 345 350  
Ile Val Met Ala Ile Val Thr Gly Ala Ile Ile Gly Lys Thr Gly Tyr  
355 360 365  
Tyr Val Pro Trp Ala Leu Ala Ser Gly Ile Leu Val Ser Ile Ser Ala  
370 375 380  
Gly Leu Val Ser Thr Phe Gln Pro Gln Thr Ser Ile Ala Ala Trp Val  
385 390 395 400  
Met Tyr Gln Phe Leu Gly Gly Val Gly Arg Gly Cys Gly Met Gln Thr  
405 410 415

Pro Val Val Ala Ile Gln Asn Ala Leu Pro Pro Gln Thr Ser Pro Ile  
420 425 430  
Gly Ile Ser Leu Ala Met Phe Gly Gln Thr Phe Gly Gly Ser Leu Phe  
435 440 445  
Leu Thr Leu Thr Glu Leu Val Phe Ser Asn Gly Leu Asp Ser Gly Leu  
450 455 460  
Arg Gln Tyr Ala Pro Thr Leu Asn Ala Gln Glu Val Thr Ala Ala Gly  
465 470 475 480  
Ala Thr Gly Phe Arg Gln Val Val Pro Ala Pro Leu Ile Ser Arg Val  
485 490 495  
Leu Leu Ala Tyr Ser Lys Gly Val Asp His Ala Phe Tyr Val Ala Val  
500 505 510  
Gly Ala Ser Gly Ala Thr Phe Ile Phe Ala Trp Gly Met Gly Arg Leu  
515 520 525  
Ala Trp Arg Gly Trp Arg Met Gln Glu Lys Gly Arg Ser Glu  
530 535 540

<210> 10  
<211> 2532  
<212> PRT  
<213> Aspergillus terreus

<400> 10  
Met Thr Pro Leu Asp Ala Pro Gly Ala Pro Ala Pro Ile Ala Met Val  
1 10 15  
Gly Met Gly Cys Arg Phe Gly Gly Ala Thr Asp Pro Gln Lys Leu  
20 25 30  
Trp Lys Leu Leu Glu Glu Gly Gly Ser Ala Trp Ser Lys Ile Pro Pro  
35 40 45  
Ser Arg Phe Asn Val Gly Gly Val Tyr His Pro Asn Gly Gln Arg Val  
50 55 60  
Gly Ser Met His Val Arg Gly Gly His Phe Leu Asp Glu Asp Pro Ala  
65 70 75 80  
Leu Phe Asp Ala Ser Phe Phe Asn Met Ser Thr Glu Val Ala Ser Cys  
85 90 95  
Met Asp Pro Gln Tyr Arg Leu Ile Leu Glu Val Val Tyr Glu Ala Leu  
100 105 110  
Glu Ala Ala Gly Ile Pro Leu Glu Gln Val Ser Gly Ser Lys Thr Gly  
115 120 125  
Val Phe Ala Gly Thr Met Tyr His Asp Tyr Gln Gly Ser Phe Gln Arg  
130 135 140  
Gln Pro Glu Ala Leu Pro Arg Tyr Phe Ile Thr Gly Asn Ala Gly Thr  
145 150 155 160  
Met Leu Ala Asn Arg Val Ser His Phe Tyr Asp Leu Arg Gly Pro Ser  
165 170 175  
Val Ser Ile Asp Thr Ala Cys Ser Thr Thr Leu Thr Ala Leu His Leu  
180 185 190

Ala Ile Gln Ser Leu Arg Ala Gly Glu Ser Asp Met Ala Ile Val Ala  
195 200 205  
Gly Ala Asn Leu Leu Leu Asn Pro Asp Val Phe Thr Thr Met Ser Asn  
210 215 220  
Leu Gly Phe Leu Ser Ser Asp Gly Ile Ser Tyr Ser Phe Asp Ser Arg  
225 230 235 240  
Ala Asp Gly Tyr Gly Arg Gly Glu Gly Val Ala Ala Ile Val Leu Lys  
245 250 255  
Thr Leu Pro Asp Ala Val Arg Asp Gly Asp Pro Ile Arg Leu Ile Val  
260 265 270  
Arg Glu Thr Ala Ile Asn Gln Asp Gly Arg Thr Pro Ala Ile Ser Thr  
275 280 285  
Pro Ser Gly Glu Ala Gln Glu Cys Leu Ile Gln Asp Cys Tyr Gln Lys  
290 295 300  
Ala Gln Leu Asp Pro Lys Gln Thr Ser Tyr Val Glu Ala His Gly Thr  
305 310 315 320  
Gly Thr Arg Ala Gly Asp Pro Leu Glu Leu Ala Val Ile Ser Ala Ala  
325 330 335  
Phe Pro Gly Gln Gln Ile Gln Val Gly Ser Val Lys Ala Asn Ile Gly  
340 345 350  
His Thr Glu Ala Val Ser Gly Leu Ala Ser Leu Ile Lys Val Ala Leu  
355 360 365  
Ala Val Glu Lys Gly Val Ile Pro Pro Asn Ala Arg Phe Leu Gln Pro  
370 375 380  
Ser Lys Lys Leu Leu Lys Asp Thr His Ile Gln Ile Pro Leu Cys Ser  
385 390 395 400  
Gln Ser Trp Ile Pro Thr Asp Gly Val Arg Arg Ala Ser Ile Asn Asn  
405 410 415  
Phe Gly Phe Gly Ala Asn Ala His Ala Ile Val Glu Gln Tyr Gly  
420 425 430  
Pro Phe Ala Glu Thr Ser Ile Cys Pro Pro Asn Gly Tyr Ser Gly Asn  
435 440 445  
Tyr Asp Gly Asn Leu Gly Thr Asp Gln Ala His Ile Tyr Val Leu Ser  
450 455 460  
Ala Lys Asp Glu Asn Ser Cys Met Arg Met Val Ser Arg Leu Cys Asp  
465 470 475 480  
Tyr Ala Thr His Ala Arg Pro Ala Asp Asp Leu Gln Leu Ala Asn  
485 490 495  
Ile Ala Tyr Thr Leu Gly Ser Arg Arg Ser Asn Phe Arg Trp Lys Ala  
500 505 510  
Val Cys Thr Ala His Ser Leu Thr Gly Leu Ala Gln Asn Leu Ala Gly  
515 520 525  
Glu Gly Met Arg Pro Ser Lys Ser Ala Asp Gln Val Arg Leu Gly Trp  
530 535 540

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Val Phe Thr Gly Gln Ala Gln Trp Phe Ala Met Gly Arg Glu Leu 545 550 555 560  
 Ile Glu Met Tyr Pro Val Phe Lys Glu Ala Leu Leu Glu Cys Asp Gly 565 570 575  
 Tyr Ile Lys Glu Met Gly Ser Thr Trp Ser Ile Ile Glu Glu Leu Ser 580 585 590  
 Arg Pro Glu Thr Glu Ser Arg Val Asp Gln Ala Glu Phe Ser Leu Pro 595 600 605  
 Leu Ser Thr Ala Leu Gln Ile Ala Leu Val Arg Leu Leu Trp Ser Trp 610 615 620  
 Asn Ile Gln Pro Val Ala Val Thr Ser His Ser Ser Gly Glu Ala Ala 625 630 635  
 Ala Ala Tyr Ala Ile Gly Ala Leu Thr Ala Arg Ser Ala Ile Gly Ile 645 650 655  
 Ser Tyr Ile Arg Gly Ala Leu Thr Ala Arg Asp Arg Leu Ala Ser Val 660 665 670  
 His Lys Gly Gly Met Leu Ala Val Gly Leu Ser Arg Ser Glu Val Gly 675 680 685  
 Ile Tyr Ile Arg Gln Val Pro Leu Gln Ser Glu Glu Cys Leu Val Val 690 695 700  
 Gly Cys Val Asn Ser Pro Ser Ser Val Thr Val Ser Gly Asp Leu Ser 705 710 715  
 Ala Ile Ala Lys Leu Glu Glu Leu Leu His Ala Asp Arg Ile Phe Ala 720 725 730  
 Arg Arg Leu Lys Val Thr Gln Ala Phe His Ser Ser His Met Asn Ser 740 745 750  
 Met Thr Asp Ala Phe Arg Ala Gly Leu Thr Glu Leu Phe Gly Ala Asp 755 760 765  
 Pro Ser Asp Ala Ala Asn Ala Ser Lys Asp Val Ile Tyr Ala Ser Pro 770 775 780  
 Arg Thr Gly Ala Arg Leu His Asp Met Asn Arg Leu Arg Asp Pro Ile 785 790 795  
 His Trp Val Glu Cys Met Leu His Pro Val Glu Phe Glu Ser Ala Phe 800 805 810  
 Arg Arg Met Cys Leu Asp Glu Asn Asp His Met Pro Lys Val Asp Arg 820 825 830  
 Val Ile Glu Ile Gly Pro His Gly Ala Leu Gly Gly Pro Ile Lys Gln 835 840 845  
 Ile Met Gln Leu Pro Glu Leu Ala Thr Cys Asp Ile Pro Tyr Leu Ser 850 855 860  
 Cys Leu Ser Arg Gly Lys Ser Ser Leu Ser Thr Leu Arg Leu Leu Ala 865 870 875  
 Ser Glu Leu Ile Arg Ala Gly Phe Pro Val Asp Leu Asn Ala Ile Asn 880 885 890 895

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Phe Pro Arg Gly Cys Glu Ala Ala Arg Val Gln Val Leu Ser Asp Leu 900 905 910  
 Pro Pro Tyr Pro Trp Asn His Glu Thr Arg Tyr Trp Lys Glu Pro Arg 915 920 925  
 Ile Ser Gln Ser Ala Arg Gln Arg Lys Gly Pro Val His Asp Leu Ile 930 935 940  
 Gly Leu Gln Glu Pro Leu Asn Leu Pro Leu Ala Arg Ser Trp His Asn 945 950 955  
 Val Leu Arg Val Ser Asp Leu Pro Trp Leu Arg Asp His Val Val Gly 960 965 970  
 Ser His Ile Val Phe Pro Gly Ala Gly Phe Val Cys Met Ala Val Met 980 985 990  
 Gly Ile Ser Thr Leu Cys Ser Ser Asp His Glu Ser Asp Asp Ile Ser 995 1000 1005  
 Tyr Ile Leu Arg Asp Val Asn Phe Ala Gln Ala Leu Ile Leu Pro Ala 1010 1015 1020  
 Asp Gly Glu Glu Gly Ile Asp Leu Arg Leu Thr Ile Cys Ala Pro Asp 1025 1030 1035  
 Gln Ser Leu Gly Ser Gln Asp Trp Gln Arg Phe Leu Val His Ser Ile 1040 1045 1050  
 Thr Ala Asp Lys Asn Asp Trp Thr Glu His Cys Thr Gly Leu Val Arg 1055 1060 1065  
 Ala Glu Met Asp Gln Pro Pro Ser Ser Leu Ser Asn Gln Gln Arg Ile 1070 1075 1080  
 Asp Arg Pro Trp Ser Arg Lys Thr Ala Pro Gln Glu Leu Trp Asp 1085 1090 1095  
 Ser Leu His Arg Val Gly Ile Arg His Gly Pro Phe Phe Arg Asn Ile 1100 1105 1110  
 Thr Cys Ile Glu Ser Asp Gly Arg Gly Ser Trp Cys Thr Phe Ala Ile 1115 1120 1125  
 Ala Asp Thr Ala Ser Ala Met Pro His Ala Tyr Glu Ser Gln His Ile 1130 1135 1140  
 Val His Pro Thr Thr Leu Asp Ser Ala Val Gln Ala Ala Tyr Thr Thr 1145 1150 1155  
 Leu Pro Phe Ala Gly Ser Arg Ile Lys Ser Ala Met Val Pro Ala Arg 1160 1165 1170  
 Val Gly Cys Met Lys Ile Ser Ser Arg Leu Ala Asp Leu Glu Ala Arg 1175 1180 1185  
 Asp Met Leu Arg Ala Gln Ala Lys Met His Ser Gln Ser Pro Ser Ala 1190 1195 1200  
 Leu Val Thr Asp Val Ala Val Phe Asp Glu Ala Asp Pro Val Gly Gly 1205 1210 1215  
 Pro Val Met Glu Leu Glu Gly Leu Val Phe Gln Ser Leu Gly Ala Ser 1220 1225 1230 1235 1240 1245

Leu Gly Thr Ser Asp Arg Asp Ser Thr Asp Pro Gly Asn Thr Cys Ser  
 1250 1255 1260  
 Ser Trp His Trp Ala Pro Asp Ile Ser Leu Val Asn Pro Gly Trp Leu  
 1265 1270 1275 1280  
 Glu Lys Thr Leu Gly Thr Gly Ile Gln Glu His Glu Ile Ser Leu Ile  
 1285 1290 1295  
 Leu Glu Leu Arg Arg Cys Ser Val His Phe Ile Gln Glu Ala Met Glu  
 1300 1305 1310  
 Ser Leu Ser Val Gly Asp Val Glu Arg Leu Ser Gly His Leu Ala Lys  
 1315 1320 1325  
 Phe Tyr Ala Trp Met Gln Lys Gln Leu Ala Cys Ala Gln Asn Gly Glu  
 1330 1335 1340  
 Leu Gly Pro Glu Ser Ser Trp Trp Arg Asp Ser Glu Gln Ala Arg  
 1345 1350 1355 1360  
 Cys Ser Leu Arg Ser Arg Val Val Ala Gly Ser Thr Asn Gly Glu Met  
 1365 1370 1375  
 Ile Cys Arg Leu Gly Ser Val Leu Pro Ala Ile Leu Arg Arg Glu Val  
 1380 1385 1390  
 Asp Pro Leu Glu Val Met Met Asp Gly His Leu Leu Ser Arg Tyr Tyr  
 1395 1400 1405  
 Val Asp Ala Leu Lys Trp Ser Arg Ser Asn Ala Gln Ala Ser Glu Leu  
 1410 1415 1420  
 Val Arg Leu Cys Cys His Lys Asn Pro Arg Ala Arg Ile Leu Glu Ile  
 1425 1430 1435 1440  
 Gly Gly Gly Thr Gly Cys Thr Gln Leu Val Val Asp Ser Leu Gly  
 1445 1450 1455  
 Pro Asn Pro Val Gly Arg Tyr Asp Phe Thr Asp Val Ser Ala Gly  
 1460 1465 1470  
 Phe Phe Glu Ala Ala Arg Lys Arg Phe Ala Gly Trp Gln Asn Val Met  
 1475 1480 1485  
 Asp Phe Arg Lys Leu Asp Ile Glu Asp Asp Pro Glu Ala Gln Gly Phe  
 1490 1495 1500  
 Val Cys Gly Ser Tyr Asp Val Val Leu Ala Cys Gln Val Leu His Ala  
 1505 1510 1515 1520  
 Thr Ser Asn Met Gln Arg Thr Leu Thr Asn Val Arg Lys Leu Lys  
 1525 1530 1535  
 Pro Gly Gly Lys Leu Ile Leu Val Glu Thr Thr Arg Asp Glu Leu Asp  
 1540 1545 1550  
 Leu Phe Phe Thr Phe Gly Leu Leu Pro Gly Trp Trp Leu Ser Glu Glu  
 1555 1560 1565  
 Pro Glu Arg Gln Ser Thr Pro Ser Leu Ser Pro Thr Met Trp Arg Ser  
 1570 1575 1580  
 Met Leu His Thr Thr Gly Phe Asn Gly Val Glu Val Glu Ala Arg Asp  
 1585 1590 1595 1600

Cys Asp Ser His Glu Phe Tyr Met Ile Ser Thr Met Met Ser Thr Ala  
 1605 1610 1615  
 Val Gln Ala Thr Pro Met Ser Cys Ser Val Lys Leu Pro Glu Val Leu  
 1620 1625 1630  
 Leu Val Tyr Val Asp Ser Ser Thr Pro Met Ser Trp Ile Ser Asp Leu  
 1635 1640 1645  
 Gln Gly Glu Ile Arg Gly Arg Asn Cys Ser Val Thr Ser Leu Gln Ala  
 1650 1655 1660  
 Leu Arg Gln Val Pro Pro Thr Glu Gly Gln Ile Cys Val Phe Leu Gly  
 1665 1670 1675 1680  
 Glu Val Glu His Ser Met Leu Gly Ser Val Thr Asn Asp Asp Phe Thr  
 1685 1690 1695  
 Leu Leu Thr Ser Met Leu Gln Leu Ala Gly Gly Thr Leu Trp Val Thr  
 1700 1705 1710  
 Gln Gly Ala Thr Met Lys Ser Asp Asp Pro Leu Lys Ala Leu His Leu  
 1715 1720 1725  
 Gly Leu Leu Arg Thr Met Arg Asn Glu Ser His Gly Lys Arg Phe Val  
 1730 1735 1740  
 Ser Leu Asp Leu Asp Pro Ser Arg Asn Pro Trp Thr Gly Asp Ser Arg  
 1745 1750 1755 1760  
 Asp Ala Ile Val Ser Val Leu Asp Leu Ile Ser Met Ser Asp Glu Lys  
 1765 1770 1775  
 Glu Phe Asp Tyr Ala Glu Arg Asp Gly Val Ile His Val Pro Arg Ala  
 1780 1785 1790  
 Phe Ser Asp Ser Ile Asn Gly Gly Glu Asp Gly Tyr Ala Leu Glu  
 1795 1800 1805  
 Pro Phe Gln Asp Ser Gln His Leu Leu Arg Leu Asp Ile Gln Thr Pro  
 1810 1815 1820  
 Gly Leu Leu Asp Ser Leu His Phe Thr Lys Arg Asn Val Asp Thr Tyr  
 1825 1830 1835 1840  
 Glu Pro Asp Lys Leu Pro Asp Asp Trp Val Glu Ile Glu Pro Arg Ala  
 1845 1850 1855  
 Phe Gly Leu Asn Phe Arg Asp Ile Met Val Ala Met Gly Gln Leu Glu  
 1860 1865 1870  
 Ser Asn Val Met Gly Phe Glu Cys Ala Gly Val Val Thr Ser Leu Ser  
 1875 1880 1885  
 Glu Thr Ala Arg Thr Ile Ala Pro Gly Leu Ala Val Gly Asp Arg Val  
 1890 1895 1900  
 Cys Ala Leu Met Asn Gly His Trp Ala Ser Arg Val Thr Thr Ser Arg  
 1905 1910 1915 1920  
 Thr Asn Val Val Arg Ile Pro Glu Thr Leu Ser Phe Pro His Ala Ala  
 1925 1930 1935  
 Ser Ile Pro Leu Ala Phe Thr Thr Ala Tyr Ile Ser Leu Tyr Thr Val  
 1940 1945 1950

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Ala Arg Ile Leu Pro Gly Glu Thr Val Leu Ile His Ala Gly Ala Gly  
1355 1960 1965  
Gly Val Gly Gln Ala Ala Ile Ile Leu Ala Gln Leu Thr Gly Ala Gly  
1970 1975 1980  
Val Phe Thr Thr Ala Gly Ser Glu Thr Lys Arg Asn Leu Leu Ile Asp  
1985 1990 2000  
Lys Phe His Leu Asp Pro Asp His Val Phe Ser Ser Arg Asp Ser Ser  
2005 2010 2015  
Phe Val Asp Gly Ile Lys Thr Arg Thr Arg Gly Lys Gly Val Asp Val  
2020 2025 2030  
Val Leu Asn Ser Leu Ala Gly Pro Leu Leu Gln Lys Ser Phe Asp Cys  
2035 2040 2045  
Leu Ala Arg Phe Gly Arg Phe Val Glu Ile Gly Lys Lys Asp Leu Glu  
2050 2055 2060  
Gln Asn Ser Arg Leu Asp Met Ser Thr Phe Val Arg Asn Val Ser Phe  
2065 2070 2075  
Ser Ser Val Asp Ile Leu Tyr Trp Gln Gln Ala Lys Pro Ala Glu Ile  
2085 2090 2095  
Phe Gln Ala Met Ser Glu Val Ile Leu Leu Trp Glu Arg Thr Ala Ile  
2100 2105 2110  
Gly Leu Ile His Pro Ile Ser Glu Tyr Pro Met Ser Ala Leu Glu Lys  
2115 2120 2125  
Ala Phe Arg Thr Met Gln Ser Gly Gln His Val Gly Lys Ile Val Val  
2130 2135 2140  
Thr Val Ala Pro Asp Asp Ala Val Leu Val Arg Gln Glu Arg Met Pro  
2145 2150 2155  
Leu Phe Leu Lys Pro Asn Val Ser Tyr Leu Val Ala Gly Gly Leu Gly  
2165 2170 2175  
Gly Ile Gly Arg Arg Ile Cys Glu Trp Leu Val Asp Arg Gly Ala Arg  
2180 2185 2190  
Tyr Leu Ile Ile Leu Ser Arg Thr Ala Arg Val Asp Pro Val Val Thr  
2195 2200 2205  
Ser Leu Gln Glu Arg Gly Cys Thr Val Ser Val Gln Ala Cys Asp Val  
2210 2215 2220  
Ala Asp Glu Ser Gln Leu Glu Ala Ala Leu Gln Gln Cys Arg Ala Glu  
2225 2230 2235  
Glu Met Pro Pro Ile Arg Gly Val Ile Gln Gly Ala Met Val Leu Lys  
2245 2250 2255  
Asp Ala Leu Val Ser Gln Met Thr Ala Asp Gly Phe His Ala Ala Leu  
2260 2265 2270  
Arg Pro Lys Val Gln Gly Ser Trp Asn Leu His Arg Ile Ala Ser Asp  
2275 2280 2285  
Val Asp Phe Phe Val Met Leu Ser Ser Leu Val Gly Val Met Gly Gly  
2290 2295 2300

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Ala Gly Gln Ala Asn Tyr Ala Ala Ala Gly Ala Phe Gln Asp Ala Leu  
2305 2310 2315  
Ala Glu His Arg Met Ala His Asn Gln Pro Ala Val Thr Ile Asp Leu  
2325 2330 2335  
Gly Met Val Gln Ser Ile Gly Tyr Val Ala Glu Thr Asp Ser Ala Val  
2340 2345 2350  
Ala Glu Arg Leu Gln Arg Ile Gly Tyr Gln Pro Leu His Glu Glu  
2355 2360 2365  
Val Leu Asp Val Leu Glu Gln Ala Ile Ser Pro Val Cys Ser Pro Ala  
2370 2375 2380  
Ala Pro Thr Arg Pro Ala Val Ile Val Thr Gly Ile Asn Thr Arg Pro  
2385 2390 2395  
Gly Pro His Trp Ala His Ala Asp Trp Met Gln Glu Ala Arg Phe Ala  
2405 2410 2415  
Gly Ile Lys Tyr Arg Asp Pro Leu Arg Asp Asn His Gly Ala Leu Ser  
2420 2425 2430  
Leu Thr Pro Ala Glu Asp Asp Asp Asn Leu His Ala Arg Leu Asn Ala  
2435 2440 2445  
Ile Ser Gln Gln Glu Ser Ile Ala Val Ile Met Glu Ala Met Ser Cys  
2450 2455 2460  
Lys Leu Ile Ser Met Phe Gly Leu Thr Asp Ser Glu Met Ser Ala Thr  
2465 2470 2475  
Gln Thr Leu Ala Gly Ile Gly Val Asp Ser Leu Val Ala Ile Glu Leu  
2485 2490 2495  
Arg Asn Trp Ile Thr Ala Lys Phe Asn Val Asp Ile Ser Val Phe Glu  
2500 2505 2510  
Leu Met Glu Gly Arg Thr Ile Ala Lys Val Ala Glu Val Val Leu Gln  
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Arg Tyr Lys Ala  
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<211> 249  
<212> PRT  
<213> Aspergillus terreus  
<400> 11  
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20 25 30  
Trp Met Lys Arg Gly Tyr Ser Cys Asn Ser Val Arg Thr Asp Asp Lys  
35 40 45  
His His Leu Arg His Leu Thr Asn Ile Gly Leu Asp Thr Pro Cys  
50 55 60  
Pro Lys Ser Leu Pro Ala Ala His Ser Ala Val Ala Ser Cys Leu Thr  
65 70 75 80

Phe Val Pro Pro Asp Pro Cys Glu Asn Thr Glu Ala Leu Gln Val Ala 95  
 85  
 Trp Asp Lys Ala Cys Cys Arg Asn Pro Thr Pro Leu Phe Phe Ile Cys 110  
 100  
 Val Ser Leu Leu Phe Ser Phe Tyr Ser Leu Trp Leu Gln Arg Gly Gly 125  
 115  
 Cys Gly Arg Tyr Gly Gly Leu His Arg Val Ser Lys Val Phe Pro Lys 140  
 130  
 Val Trp Pro Asp Asp Met Asp Ser Gln Leu Pro Ser Arg Leu Gln Thr 160  
 145  
 Leu Val Ser Lys Arg Lys Pro Glu Pro Ala Pro Asn Asn Ser Thr Tyr 175  
 165  
 Ile Ser Lys Gly Tyr Ala Thr Phe Phe Asn Gln Phe Ser Leu Pro Ser 190  
 180  
 Val Asp Val Thr Gln Ile Leu Asn Gln Thr Leu Gln His His Asp Val 205  
 195  
 Glu Thr Ile Asn Leu Asp Cys Gly Ser Gly Leu Leu Thr Leu Arg Thr 220  
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 Gln Leu Arg Ile Leu Leu Ile Gly Lys Pro Lys Ile Ile Lys Pro Phe 240  
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 Ser Gly Leu Arg Thr Thr Ile Asn Glu 245  
  
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 <211> 742  
 <212> PRT  
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 <400> 12  
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 Asp Glu Cys Arg Ile Arg Lys Val Arg Cys Ser Lys Glu Gly Pro Lys 30  
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 Cys Ser His Cys Leu Arg Tyr Asn Leu Pro Cys Glu Phe Ser Asn Lys 45  
 35  
 Val Ala Arg Asp Val Glu Lys Leu Gly Ser Arg Val Gly Asp Ile Glu 60  
 50  
 His Ala Leu Gln Arg Cys Leu Ser Phe Ile Asp Ala His Gln Gly Phe 80  
 65  
 Arg Asp Leu Ser Arg Pro Gln Ser Gln Glu Ser Gly Tyr Thr Ser Ser 95  
 85  
 Thr Ser Ser Glu Glu Cys Glu Val Asn Leu Tyr Ser Gly Lys His Thr 110  
 100  
 Ser Pro Thr Glu Glu Asp Gly Phe Thr Pro Leu Leu His Gly Tyr Gly Ser 125  
 115  
 Phe Val Ser Leu Val Met Glu Ala Gln Ala Ala Asn Ala Asn Leu Thr 140  
 130

Ser Trp Leu Pro Val Asp Met Thr Ser Gly Gln Val Ala Glu Met Val 160  
 145  
 Ala Phe Asp Arg Gln Ala Val Ser Ala Val Arg Ser Lys Val Ala Glu 175  
 165  
 Ala Asn Glu Thr Leu Gln Gln Ile Ile Glu Asp Ile Pro Thr Leu Ser 190  
 180  
 Ala Ser Glu Asn Asp Thr Phe Leu Pro Ser Leu Pro Pro Arg Ala Leu 205  
 195  
 Val Glu Pro Ser Ile Asn Glu Tyr Phe Lys Lys Leu His Pro Arg Leu 220  
 210  
 Pro Ile Phe Ser Arg Gln Thr Ile Met Asp Ala Val Glu Ser Gln Tyr 240  
 225  
 Thr Ile Arg Thr Gly Pro Pro Asp Leu Val Trp Ile Thr Ser Phe Asn 255  
 245  
 Cys Ile Val Leu Gln Ala Leu Thr Gln Thr Ser Ile Ala Asn Lys Val 270  
 260  
 Val Gly Cys Thr Gly Gln Asp Ile Pro Ile Asp Tyr Met Ile Ile Ser 285  
 275  
 Leu Leu Arg Asn Ile Arg Gln Cys Tyr Asn Arg Leu Glu Thr Leu Val 300  
 290  
 Lys Pro Arg Leu Ser Asn Ile Arg Ala Leu Phe Cys Leu Ala Leu Val 320  
 305  
 Ala Met Glu Tyr Phe Asp Phe Ala Ile Phe Leu Thr Ile Phe Ala Gln 335  
 325  
 Val Cys Glu Leu Ser Arg Leu Ile Gly Leu His Leu Thr Thr Thr Thr 350  
 340  
 Pro Pro Thr Glu Asp Gly Ala Val Gly Asp Gln Pro Lys Asp Leu Phe 365  
 355  
 Trp Ser Ile Phe Leu Val Asp Lys His Val Ser Ile Ile Gly Gly Lys 380  
 370  
 Ala Cys Leu Leu Pro Ser Tyr Asp Cys Ser Val Pro Leu Pro Pro Tyr 400  
 385  
 Asp Ser Ala Ala Pro Leu Pro Asn Ala Phe Ala Ala Arg Ile Arg Leu 415  
 405  
 Ala Phe Ile Leu Glu Glu Ile Tyr Leu Gly Leu Tyr Ser Ala Lys Ser 430  
 420  
 Ser Lys Met Glu Gln Ser Arg Val Arg Arg Arg Ile Arg Arg Ile Ala 445  
 435  
 Arg Lys Leu Ser Gln Trp His Val Gln His Glu His Val Leu Arg Thr 460  
 450  
 Gly Asp Pro Asn Arg Pro Leu Glu Glu Tyr Ile Cys Ala Thr Gln Leu 480  
 465  
 Arg Phe Ala Leu Ser Ser Cys Trp Val Leu Leu His Lys Arg Ile Trp 495  
 485



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Ser Gln Glu Arg Gly Ala Val Cys Leu Gln His Ala Arg Asp Cys Leu  
500 505 510  
Met Leu Phe Lys Gln Leu Cys Asp Gly Cys Lys Ser Gly Phe Ser Asn  
515 520 525  
Phe Asp Ser Ile Val Leu Asn Tyr Ser Leu Ile Ser Phe Met Gly Ile  
530 535 540  
Tyr Val His Ile Val Glu Glu Asp Gln Pro Ile His Ser Gln Asp Met  
545 550 555  
Glu Ile Leu Thr Phe Phe Ala Ile Tyr Thr Asn Arg Ser Ala Ser Asn  
565 570 575  
Arg Ser Ser Ala Ser Ile Ser Tyr Lys Leu Ser Gln Val Ala Ser Arg  
580 585 590  
Cys Ser Asp Ile Ala Leu Leu Leu Gln Asn Leu Arg Glu Arg Phe  
595 600 605  
Ile Pro Thr Thr Ile Ser Arg Ser Pro Thr Pro Ser Trp Asn Glu Pro  
610 615 620  
Thr Tyr Met Asp Tyr Asp Val Ala Asn Ala Ser Thr Ser Thr Thr Ser  
625 630 635  
Thr Gly Ser Ser Tyr Asn Leu Asn Ile Ser Pro Leu Gly Val Pro Gly  
645 650 655  
Asp Gly Gln Val Trp Asp Ile Tyr Phe Asn Pro Arg Glu Ile Pro Met  
660 665 670  
Asp Gly Thr Ile Ala Thr Pro Ser Glu Asp Ala Thr Gln Asp Leu Leu  
675 680 685  
Ser Asn Asp Ala Gly Gln Cys Leu Gly Phe Pro Asp Phe Ser Leu Gly  
690 695 700  
Ile Asp Asn Phe Ser Asp Phe Pro Leu Gly Ile Asp Met Thr Ser Gln  
705 710 715  
Ser Glu Phe Gly Leu Ile Met Glu Glu Asp Ile Ile Arg Tyr Glu Arg  
725 730 735  
Leu Leu Asp Arg Pro Val  
740

<210> 13  
<211> 301  
<212> PRT  
<213> Aspergillus terreus  
<400> 13  
Met Glu Ser Lys Val Gln Thr Asn Val Pro Leu Pro Lys Ala Pro Leu  
1 5 10 15  
Thr Gln Lys Ala Arg Gly Lys Arg Thr Lys Gly Ile Pro Ala Leu Val  
20 25 30  
Ala Gly Ala Cys Ala Gly Ala Val Glu Ile Ser Ile Thr Tyr Pro Phe  
35 40 45  
Glu Ser Ala Lys Thr Arg Ala Gln Leu Lys Arg Arg Asn His Asp Val  
50 55 60

27

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Ala Ala Ile Lys Pro Gly Ile Arg Gly Trp Tyr Ala Gly Tyr Gly Ala  
65 70 75 80  
Thr Leu Val Gly Thr Thr Leu Lys Ala Ser Val Gln Phe Ala Ser Phe  
85 90 95  
Asn Ile Tyr Arg Ser Ala Leu Ser Gly Pro Asn Gly Glu Leu Ser Thr  
100 105 110  
Gly Ala Ser Val Leu Ala Gly Phe Gly Ala Gly Val Thr Glu Ala Val  
115 120 125  
Leu Ala Val Thr Pro Ala Glu Ala Ile Lys Thr Lys Ile Ile Asp Ala  
130 135 140  
Arg Lys Val Gly Asn Ala Glu Leu Ser Thr Thr Phe Gly Ala Ile Ala  
145 150 155 160  
Gly Ile Leu Arg Asp Arg Gly Pro Leu Gly Phe Phe Ser Ala Val Gly  
165 170 175  
Pro Thr Ile Leu Arg Gln Ser Ser Asn Ala Ala Val Lys Phe Thr Val  
180 185 190  
Tyr Asn Glu Leu Ile Gly Leu Ala Arg Lys Tyr Ser Lys Asn Gly Glu  
195 200 205  
Asp Val His Pro Leu Ala Ser Thr Leu Val Gly Ser Val Thr Gly Val  
210 215 220  
Cys Cys Ala Trp Ser Thr Gln Pro Leu Asp Val Ile Lys Thr Arg Met  
225 230 235 240  
Gln Ser Leu Gln Ala Arg Gln Leu Tyr Gly Asn Thr Phe Asn Cys Val  
245 250 255  
Lys Thr Leu Leu Arg Asn Glu Gly Ile Gly Val Phe Trp Ser Gly Val  
260 265 270  
Trp Phe Arg Thr Gly Arg Leu Ser Leu Thr Ser Ala Ile Met Phe Pro  
275 280 285  
Val Tyr Glu Lys Val Tyr Lys Phe Leu Thr Gln Pro Asn  
290 295 300

<210> 14  
<211> 490  
<212> PRT  
<213> Aspergillus terreus  
<400> 14  
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Glu Ile Cys His Trp Ala Ser Asn Leu Ala Thr Asp Asp Ile Pro Pro  
20 25 30  
Asp Val Leu Leu Arg Ala Lys Tyr Leu Ile Leu Asp Gly Ile Ala Cys  
35 40 45  
Ala Trp Val Gly Ala Arg Val Pro Trp Ser Glu Lys Tyr Val Gln Ala  
50 55 60  
Thr Met Ser Phe Glu Pro Pro Gly Ala Cys Arg Val Ile Gly Tyr Gly  
65 70 75 80

28

Gln Lys Leu Gly Pro Val Ala Ala Met Thr Asn Ser Ala Phe Ile  
85 90  
Gln Ala Thr Glu Leu Asp Asp Tyr His Ser Glu Ala Pro Leu His Ser  
100 110  
Ala Ser Ile Val Leu Pro Ala Val Phe Ala Ala Ser Glu Val Leu Ala  
115 125  
Glu Gln Gly Lys Thr Ile Ser Gly Ile Ala Val Ile Leu Ala Ala Ile  
130 135 140  
Val Gly Phe Glu Ser Gly Pro Arg Ile Gly Lys Ala Ile Tyr Gly Ser  
145 155  
Asp Leu Leu Asn Asn Gly Trp His Cys Gly Ala Val Tyr Gly Ala Pro  
165 170 175  
Ala Gly Ala Leu Ala Thr Gly Lys Leu Leu Gly Thr Pro Asp Ser  
180 185 190  
Met Glu Asp Ala Leu Gly Ile Ala Cys Thr Gln Ala Cys Gly Leu Met  
195 200 205  
Ser Ala Gln Tyr Gly Gly Met Val Lys Arg Val Gln His Gly Phe Ala  
210 215 220  
Ala Arg Asn Gly Leu Leu Gly Gly Leu Ala His Gly Gly Tyr Glu  
225 230 235  
Ala Met Lys Gly Val Leu Glu Arg Ser Tyr Gly Gly Phe Leu Lys Met  
245 250 255  
Phe Thr Lys Gly Asn Gly Arg Glu Pro Pro Tyr Lys Glu Glu Glu Val  
260 265 270  
Val Ala Gly Leu Gly Ser Phe Thr His Thr Phe Thr Ile Arg Ile Lys  
275 280 285  
Leu Tyr Ala Cys Cys Gly Leu Val His Gly Pro Val Glu Ala Ile Glu  
290 295 300  
Asn Leu Gln Arg Arg Tyr Pro Glu Leu Leu Asn Arg Ala Asn Leu Ser  
305 310 315 320  
Asn Ile Arg His Val His Val Gln Leu Ser Thr Ala Ser Asn Ser His  
325 330 335  
Cys Gly Trp Ile Pro Glu Glu Arg Pro Ile Ser Ser Ile Ala Gly Gln  
340 345 350  
Met Ser Val Ala Tyr Ile Leu Ala Val Gln Leu Val Asp Gln Gln Cys  
355 360 365  
Leu Leu Ala Gln Phe Ser Glu Phe Asp Asp Asn Leu Glu Arg Pro Glu  
370 375 380  
Val Trp Asp Leu Ala Arg Lys Val Thr Pro Ser His Ser Glu Glu Phe  
385 390 395 400  
Asp Gln Asp Gly Asn Cys Leu Ser Ala Gly Arg Val Arg Ile Glu Phe  
405 410 415  
Asn Asp Gly Ser Ser Val Thr Glu Thr Val Glu Lys Pro Leu Gly Val  
420 425 430

Lys Glu Pro Met Pro Asn Glu Arg Ile Leu His Lys Tyr Arg Thr Leu  
435 440 445  
Ala Gly Ser Val Thr Asp Glu Thr Arg Val Lys Glu Ile Glu Asp Leu  
450 455 460  
Val Leu Ser Leu Asp Arg Leu Thr Asp Ile Ser Pro Leu Leu Glu Leu  
465 470 475 480  
Leu Asn Cys Pro Val Lys Ser Pro Leu Val  
485 490  
<210> 15  
<211> 488  
<212> PRT  
<213> Aspergillus terreus  
<400> 15  
Met Gly Arg Gly Asp Thr Glu Ser Pro Asn Pro Ala Thr Thr Ser Glu  
1 5 10 15  
Gly Ser Gly Gln Asn Glu Pro Glu Lys Lys Gly Arg Asp Ile Pro Leu  
20 25 30  
Trp Arg Lys Cys Val Ile Thr Phe Val Val Ser Trp Met Thr Leu Val  
35 40 45  
Val Thr Phe Ser Ser Thr Cys Leu Leu Pro Ala Ala Pro Glu Ile Ala  
50 55 60  
Asn Glu Phe Asp Met Thr Val Glu Thr Ile Asn Ile Ser Asn Ala Gly  
65 70 75 80  
Val Leu Val Ala Met Gly Tyr Ser Ser Leu Ile Trp Gly Pro Met Asn  
85 90 95  
Lys Leu Val Gly Arg Arg Thr Ser Tyr Asn Leu Ala Ile Ser Met Leu  
100 105 110  
Cys Ala Cys Ser Ala Gly Thr Ala Ala Ala Ile Asn Glu Lys Met Phe  
115 120 125  
Ile Ala Phe Arg Val Leu Ser Gly Leu Thr Gly Thr Ser Phe Met Val  
130 135 140  
Ser Gly Gln Thr Val Leu Ala Asp Ile Phe Glu Pro Val Tyr Arg Gly  
145 150 155 160  
Thr Ala Val Gly Phe Met Ala Gly Thr Leu Ser Gly Pro Ala Ile  
165 170 175  
Gly Pro Cys Val Gly Gly Val Ile Val Thr Phe Thr Ser Trp Arg Val  
180 185 190  
Ile Phe Trp Leu Gln Leu Gly Met Ser Gly Leu Gly Leu Val Leu Ser  
195 200 205  
Leu Leu Phe Phe Pro Lys Ile Glu Gly Thr Ser Glu Lys Val Ser Thr  
210 215 220  
Ala Phe Lys Pro Thr Thr Leu Val Ser Ile Ile Ser Lys Phe Ser Pro  
225 230 235 240  
Thr Asp Val Leu Lys Gln Trp Val Tyr Pro Asn Val Phe Leu Ala Val  
245 250 255

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Ser Ala Trp Glu Ile Cys Pro Leu His Leu Leu Glu Thr Lys Cys Ser  
260 265 270  
Cys Arg Lys Gln Lys Asp Leu Cys Cys Gly Leu Leu Ala Ile Thr Gln  
275 280 285  
Tyr Ser Ile Leu Thr Ser Ala Arg Ala Ile Phe Asn Ser Arg Phe His  
290 295 300  
Leu Thr Thr Ala Leu Val Ser Gly Leu Phe Tyr Leu Ala Pro Gly Ala  
305 310 315  
Gly Phe Leu Ile Gly Ser Leu Val Gly Gly Lys Leu Ser Asp Arg Thr  
320 325 330 335  
Val Arg Arg Tyr Ile Val Lys Arg Gly Phe Arg Leu Pro Gln Asp Arg  
340 345 350  
Leu His Ser Gly Leu Ile Thr Leu Phe Ala Val Leu Pro Ala Gly Thr  
355 360 365  
Leu Ile Tyr Gly Trp Thr Leu Gln Glu Asp Lys Gly Gly Met Val Val  
370 375 380  
Pro Ile Ile Ala Ala Phe Phe Ala Gly Trp Gly Leu Met Gly Ser Phe  
385 390 395  
Asn Cys Leu Asn Thr Tyr Val Ala Val Glu Ala Leu Pro Arg Asn Arg  
400 405 410 415  
Ser Ala Val Ile Ala Gly Lys Tyr Met Ile Gln Tyr Ser Phe Ser Ala  
420 425 430  
Gly Ser Ser Ala Leu Val Val Pro Val Ile Asp Ala Leu Gly Val Gly  
435 440 445  
Trp Thr Phe Thr Leu Cys Val Val Ala Ser Thr Ile Ala Gly Leu Ile  
450 455 460  
Thr Ala Ala Ile Ala Arg Trp Gly Ile Asn Met Gln Arg Trp Ala Glu  
465 470 475 480  
Arg Ala Phe Asn Leu Pro Thr Gln  
485  
<210> 16  
<211> 316  
<212> PRT  
<213> Aspergillus terreus  
<400> 16  
Met Thr Leu Gln Ile Ile Val Ile Ala Ala Thr Ala Val Ile Tyr Phe  
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Leu Thr Arg Tyr Phe Asn Arg Thr Asp Ile Pro Lys Ile Lys Gly Ile  
20 25 30  
Pro Glu Ile Pro Gly Val Pro Ile Phe Gly Asn Leu Ile Gln Leu Gly  
35 40 45  
Val Lys His Ala Thr Val Ala Arg Lys Trp Ser Ser Glu Phe Gly Pro  
50 55 60  
Val Phe Gln Ala Arg Leu Glu Asn Arg Arg Val Ile Phe Ala Asn Thr  
65 70 75 80

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Phe Glu Ser Thr Arg Gln Leu Trp Ile Lys Glu Gln Ser Ser Met Ile  
85 90 95  
Ser Arg Pro Thr Phe His Thr Phe His Gly Val Val Ser Ser Ser Gln  
100 105 110  
Gly Phe Thr Ile Gly Thr Ser Pro Trp Asp Glu Ser Cys Lys Arg Arg  
115 120 125  
Arg Lys Ala Ala Thr Ala Leu Asn Arg Val Ala Val Gln Ser Tyr  
130 135 140  
Met Pro Ile Ile Asp Leu Glu Ser Met Ala Ser Ile Lys Glu Leu Leu  
145 150 155 160  
Lys Asp Ser Gln Gly Lys Ile Asp Ile Asn Pro Thr Pro Tyr Phe  
165 170 175  
Gln Arg Phe Ala Leu Asn Thr Ser Leu Thr Leu Asn Tyr Gly Tyr Arg  
180 185 190  
Ile Glu Gly Asn Val Asn Asp Gln Leu Leu Arg Arg Ile Cys Glu Val  
195 200 205  
Gln Arg Gly Val Ala Asn Leu Arg Ser Thr Ser Asn Asn Trp Gln Asp  
210 215 220  
Tyr Val Pro Leu Leu Arg Leu Phe Ser Asn Arg Ser Asn Gln Ala Lys  
225 230 235 240  
His Leu Arg Ala Arg Arg Asp Lys Tyr Met Ala Phe Leu Phe Asp Ile  
245 250 255  
Leu Lys Asp Arg Met Ala Lys Gly Thr Asp Lys Pro Cys Ile Thr Gly  
260 265 270  
Asn Ile Leu Lys Asn Pro Glu Thr Lys Leu Thr Asp Ala Glu Ile Lys  
275 280 285  
Ser Ile Cys Leu Thr Met Val Ser Ala Gly Leu Asp Thr Val Pro Gly  
290 295 300  
Asn Leu Ile Met Gly Ile Ala Tyr Leu Ser Ser Glu Asp Gly Gln Arg  
305 310 315 320  
Ile Gln Gln Lys Ala Tyr Glu Glu Ile Met Ser Val Tyr Pro Asn Gly  
325 330 335  
Asp Ala Trp Glu Arg Cys Leu Val Glu Glu Lys Val Pro Tyr Ile Thr  
340 345 350  
Ala Leu Val Lys Glu Thr Leu Arg Phe Trp Thr Val Met Pro Ile Cys  
355 360 365  
Ile Pro Arg Val Asn Ile Lys Glu Val Ile Tyr Asn Gly Ala Arg Ile  
370 375 380  
Pro Ala Gly Thr Thr Phe Phe Met Asn Ala Trp Ala Ala Asn Tyr Asp  
385 390 395 400  
Glu Asp His Phe Asp Met Pro Asn Arg Phe Leu Pro Glu Arg Tyr Leu  
405 410 415  
Glu Pro Ser Glu Gly Phe Gly Thr Pro His Tyr Ser Phe Gly Ala Gly  
420 425 430

Thr Arg Met Cys Ala Ala Ser His Leu Ala Ser Arg Glu Leu Tyr Thr  
435 445  
Val Phe Leu Arg Phe Ile Val Ala Phe Thr Ile Glu Pro Ala Gln Asn  
450 460  
Pro Ala Asp Met Pro Val Leu Asp Ala Ile Glu Cys Asn Ala Thr Pro  
465 470 475 480  
Thr Ser Met Thr Thr Glu Pro Lys Pro Phe Lys Val Gly Phe Lys Pro  
485 490 495  
Arg Asp Glu Thr Ser Leu Arg Arg Trp Ile Ala Glu Ser Glu Glu Arg  
500 510  
Thr Lys Glu Leu  
515

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<211> 481  
<212> PRT  
<213> Aspergillus terreus

<400> 17  
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Ser Ala Ser Thr Leu Asn Gly Lys Leu Thr Leu Ser Glu Thr Lys Val  
20 25 30

Thr Gly Ala Val Gln Leu Ala Cys Thr Asn Ser Pro Pro Asp Ile Tyr  
35 40 45

Ile Asp Pro Asp Asp Ser Val Ser Val Arg Ala Ala His Asp Leu  
50 55 60

Ala Leu Asp Phe Gly Arg Val Phe Gly Lys Asn Ala Thr Val Arg Phe  
65 70 75 80

Thr Asn Glu Thr His Pro Thr Ser Met Ala Ile Ile Ala Gly Thr Ile  
85 90 95

Asp Lys Ser Thr Phe Leu Gln Arg Leu Ile Ala Asp His Lys Leu Asp  
100 105 110

Val Thr Ser Ile Arg Gly Gln Trp Glu Ser Tyr Ser Ser Ala Leu Val  
115 120 125

Leu Gly Pro Ala Lys Gly Ile Gln Asn Ala Leu Val Ile Ala Gly Ser  
130 135 140

Asp Arg Arg Gly Ala Ile Tyr Gly Leu Tyr Asp Ile Ser Glu Gln Ile  
145 150 155 160

Gly Val Ser Pro Leu Phe Thr Trp Thr Asp Val Thr Pro Thr Lys Leu  
165 170 175

Asp Ala Ile Tyr Ala Leu Asp Val Gln Lys Val Gln Gly Pro Pro Ser  
180 185 190

Val Lys Tyr Arg Gly Ile Phe Ile Asn Asp Glu Ala Pro Ala Leu His  
195 200 205

Asn Trp Ile Leu Ala Asn Tyr Gly Glu Val Glu Asn Gly Asp Pro Ala  
210 215 220

Phe Ile Ser Arg Phe Tyr Ala His Val Phe Glu Leu Ile Leu Arg Leu  
225 230 235 240  
Lys Gly Asn Tyr Leu Trp Pro Ala Met Trp Ser Asn Met Phe Tyr Val  
245 250 255

Asp Asp Thr Asn Asn Gly Pro Leu Ala Asp Tyr Tyr Gly Val Val Met  
260 265 270

Gly Thr Ser His Thr Gly Met Thr Val Gly Thr Pro Cys Leu Lys Ala  
275 280 285

His Ala Asp Tyr Glu Lys Glu Pro Met Ala Arg Ala Thr Asn Glu Gln  
290 295 300

Ser Gln Phe Leu Asn Gly Thr Trp Asp Trp Ile Ser Asn Glu Val Asn  
305 310 315 320

Val Lys Ala Phe Met Arg Glu Gly Val Ile Arg Ser Gln His Trp Glu  
325 330 335

Thr Ala Tyr Thr Met Gly Met Arg Gly Leu Gly Asp Ala Ala Ser Pro  
340 345 350

Thr Leu Asn Ala Thr Val Glu Glu Ser Ile Val Ser Trp Gln Glu Ser  
355 360 365

Val Leu Ser Asp Ile Leu Asn Lys Thr Asn Leu Ser Asn Val Val Gln  
370 375 380

Pro Phe Val Leu Phe Asp Glu Leu Gly Thr Tyr Tyr Glu Ser Gly Met  
385 390 395 400

Thr Val Pro Asp Gln Val Thr Leu Ile Tyr Pro Asp Asp Asn Ala Gly  
405 410 415

Asn Met Leu Arg Leu Pro Leu Gln Asn Glu Thr Gly Arg Ser Gly Gly  
420 425 430

Ala Gly Ile Tyr Tyr His Phe Asp Met Asn Ala Pro Pro Arg Cys Tyr  
435 440 445

Lys Trp Ile Asn Thr Ala Gln Leu Ile Arg Thr Trp Asp Gln Leu Arg  
450 455 460

Ala Ala Tyr Ser His Gly Ala Gln Thr Val Trp Val Ala Asn Ile Gly  
465 470 475 480

Asp  
<210> 18  
<211> 33000  
<212> DNA  
<213> Aspergillus terreus

<400> 18  
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aattaggaa actgctggc tatctatcc agagcttag ctaggtagt tacctgact 120  
gtttgcga gctgtaggc ctgagggcg actaatatt aggtatttt tacaagccg 180  
gaatagagc ttatagatc atagagctt agaagatct actgctaat atctattct 240  
aaatacaa gaattctaat agagcttct taaagatc ctctcttaag agtatgctc 300  
agtaagaa ttcaaatat taacagcct taataagca gtctcttag ttgctgctc 360  
cgttagagc gctcaatag tttaagctt tagcttaga ttaggagat aactagctc 420  
cgttagaca ttcttctta tttaagcgc gtaactagg cagtattag ctagtggcg 480  
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<210> 22

<211> 872

<212> DNA

<213> Aspergillus terreus

<400> 22

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ctctcagc caattacca gaagcact gggaaagag agtctcgc gcaaaaggc 240  
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tggtagcgc cagcgggc gcaatgac cagtgatg ctgtatca tgaacctcc 360  
gactcgta tgggtgggt gttggagtc gttgacac ccatctct tgaacctcc 420  
cgtatgga tccggaggt catggccaa ggggttga cactgctc gctgacaa 480  
ctcaactg tgcagtgt tctcaagg tcaagcgc tcaagcct gaaatgac 540  
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gaaagctg tgcgtgag tgcagcgc atgagacc ccaggtcg gcagagcg 660  
gaaatagc atcttacc gatatgag ctgcagag cccggtaa acggttct 720  
gccaaatg agaaccaa cggggacc atcggttg gttggatc acggttct 780  
ccggccgt tcttgctc taaggacc agatgatg tagctact gcgtatca 840  
tcagctga agtgttcc cgaagacg ttgagtae ttgagact ttcagctg 900  
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<210> 23

<211> 771

<212> DNA

<213> Aspergillus terreus

<400> 23

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gatttgcg tgcagact gaagatgct gttggcgc attctccc ccgagagcg 660  
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<210> 24

<211> 1253

<212> DNA

<213> Aspergillus terreus

<400> 24

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cgtggctc acaagcag gttatacaa gatccgctg cagtcacga tataacca 66c  
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<210> 25

<211> 1394

<212> DNA

<213> Aspergillus terreus

<400> 25

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acagggac atactcagc aattcaat atacgctg ctccgggt cgaagctg 240  
gacggaga gtgcattcg ctgcgcgc taccatag caccctgc cgtctgca 300  
gtgcacaa gtgtctac acatcag ccttcaatg catgagcg gttctgtg 360  
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ggttgaga cgggaaac gcaagatgc gagaagctc gggaaagc acgtcgag 480  
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cgcgcctg cagggacc accgctgc ggtatcag cccaactg gactggcg 660  
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<210> 26

<211> 3504

<212> DNA

<213> Aspergillus terreus

<400> 26

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ggtctgccc cctggcca gaaacccc ttgaagtc agtcttct cttgcaag 240  
cgactgta gctggaga gctagcga tggaaatgc agatagca cgcactaa 300  
gtcggagg atggcagat atgagatc tctcgttc catgttgc agaatctg 360  
tgcttacc gacccccc ttgtactc tcaattgc tgaattac tgggtctg 420  
tcactgca tctaccgg ggtctgct accagatc cctcttca tcaaacgc 480  
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tcaaatat cctcggaa aagaagat atagttgag actgagtt cctcgaa 660  
aagaattc cagtcagc gaaactgc tggagctc atgctgca tctctcat 720  
gttccaca tggtagca gtaacttg tctatagg gtcagtae ctagcaga 780  
ataatgac gctctctc ttccgaga tggcaact cgtctcagc tctggtgt 840  
cagctcgtt gctcgtct ggtccttg ctttgaact cgggttga atcagact 900









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8035

<210> 30  
<211> 888  
<212> DNA  
<213> Aspergillus terreus

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taataataa taataataa atcaaaaga ggggaataac gtaataact gtcgacac 180  
acataaaca taataataa taataataa taataataa taataataa taataataa 240  
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taacaccta atctgacga gatctaaag ctaaaacc cgcgcgac ttcgagag 660  
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gtacacaga tctcaataa gacgttgag caccagag ttgagatc atccgtgat 780  
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<210> 31  
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<210> 32  
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<213> Aspergillus terreus

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